Collecting and Processing Data on Fish Eggs and Larvae in the California Current Region

DAVID KRAMER, MARY J. KALIN, ELIZABETH G. STEVENS, JAMES R. THRAILKILL, and JAMES R. ZWEIFEL
National Marine Fisheries Service, Circulars

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SEATTLE, WA

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COLLECTING AND PROCESSING DATA ON FISH EGGS AND LARVAE IN THE CALIFORNIA CURRENT REGION

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ABSTRACT

Descriptions are given for the methods used by the California Cooperative Oceanic Fisheries Investigations to collect and process plankton. These include details of the design of the station pattern in the survey area, the gear and methods used for plankton hauls, measuring plankton, and sorting plankton for fish eggs and larvae; some procedures for identifying fish eggs and larvae; details of “hand” processing data for standardization of numbers of organisms collected in all plankton hauls; calibration of flowmeters; and some new procedures for automatic data processing.

INTRODUCTION

For more than 20 years the California Cooperative Oceanic Fisheries Investigations (CalCOFI) have conducted a program of intensive research in the California Current region in a designated area of approximately 500,000 square miles from the California-Oregon border to the tip of Baja California. The investigations were originated in 1949 to determine the reason for the decline of the Pacific sardine fishery. Since then, the data have contributed a wealth of information from which it has been possible to study the effects of the biological, physical, and chemical environment on all the resources in the area.

The chief participants in CalCOFI in ships, personnel, equipment, shoreside facilities, and data collection, processing and analyses are the California Department of Fish and Game (CF&G) on the evaluation of resources by census of young and adult fishes, the University of California, Scripps Institution of Oceanography (SIO), on the studies of the physical and chemical data and selected groups of invertebrates, and the National Oceanic and Atmospheric Administration (NOAA), National Marine Fisheries Service (NMFS), formerly the Bureau of Commercial Fisheries, on the evaluation of resources by censuses of fish eggs and larvae. Other participating groups are the California Academy of Sciences and the Stanford University, Hopkins Marine Station, chiefly in laboratory research.

It is the purpose of this report to describe the methods and gear used by the CalCOFI for collecting and processing data on fish eggs and larvae. Some of these have been described in varying detail (Ahlstrom, 1948, 1956) but none with the full treatment that we feel is warranted, as now reported here, in view of the requests for greater detail by visitors to our laboratory and some investigators who have cooperated with us in data collection at sea.
THE SURVEY PATTERN AND AREA

The pattern of stations (Fig. 1) covered by most of the CalCOFI surveys was designed originally on the basis of a centric-systematic-area sampling scheme (Milne, 1959) to determine the major spawning areas of the Pacific sardine off the coasts of the United States and Baja California, Mexico. This was done by conducting surveys on lines spaced 120 miles apart from the Columbia River to Sebastian Vizcaino Bay. As the spawning areas were delimited, additional lines of stations were added between the cardinal lines, and the surveys were concentrated off the coasts of California and Baja California.

The original lines of the pattern were based on line 80 off Point Conception, Calif., and set parallel to that line 120 miles apart, north to line 10 off the United States-Canadian border, and south to line 120 off Point Eugenia, Baja California. They were plotted to extend 30° southwest of lines of latitude, thus perpendicular to the coast of central California, north of Point Conception. It was intended that the 120-mile spacing would allow for additional lines to be plotted 12 miles apart between the cardinal lines and still be designated by whole numbers without resorting to fractions. However, when lines were added between cardinal lines, it was deemed sufficient to space them at 40-mile intervals. Thus, the major pattern consists of the cardinal lines in multiples of 10 ending in 0's and the ordinal lines ending in 3's and 7's. During the course of the investigations, lines were added finally to include line 157 just south of Cape San Lucas, Baja California.

The stations on the lines were laid out on the basis of a perpendicular to line 80, at a point designated station 80.60. The perpendicular, through all lines parallel to line 80, aligned the stations designated as 60's.

Most of the original stations shoreward and seaward from station 60 were plotted 40 miles apart, which allowed stations between the 40-mile points to be plotted as close as 4 miles apart and still retain whole numbers. In most cases, stations between the 40-mile points have been only 20 miles apart. Those closer than 20 miles, e.g., inshore or near islands, were so placed simply because a 20-mile spacing would have placed a station on land and the omission of such a station would have left too large a space between the last plotted station and the land. Closer spacing than 4 miles, using fractions for station numbers, has been resorted to in some instances, e.g., to locate the eggs of the increasingly rare Pacific sardine during the peak of its spawning season.

DATA COLLECTION

The Research Vessel

A vessel used to collect oceanographic data should be a relatively stable platform at sea and be capable of reasonably rapid coverage of large areas for long periods of time. Today, most vessels used for biological and hydrographic research on the high seas are 100 or more feet long, powered to cruise at 10 to 15 knots, and capable of staying at sea for 14 to 30 days. Some ships are of multipurpose design in that they can collect plankton and hydrographic data and convert to fishing operations to handle large fishing nets, trawls, seines, etc.

Gear common to the collections of plankton and hydrographic data are power winches equipped with high-strain wire, an over-the-side platform for handling gear clear of the ship's side, and a weight at the end of the wire. Shipboard facilities should include sheltered spaces for laboratory work and adequate storage spaces for gear and samples.

Essential gear for the collection of plankton are fine mesh plankton nets with detachable cod ends, rings to keep net mouths open, a bridle assembly to tow the nets, a cable clamp to attach a lead line to the towing cable, an inclinometer to measure angles of stray during a net tow, and flowmeters to measure water volume strained by the nets.

Some of the gear described here have specifications included in figure captions for which simple descriptions will suffice to allow duplication without need for further details. Some are purchased from their manufacturers or distributors. Others have specifications too detailed to be included here. Table 1 lists all of the gear and materials described, the figures in which they are illustrated, and the places of their descriptions, and or sources of specifications and suppliers.

The winches used for plankton tows or hydrographic casts should be electric or hydraulic and
Figure 1.—Basic station plan of the California Cooperative Oceanic Fisheries Investigations (CalCOFI) since 1950. Stations in the Gulf of California were occupied only on special surveys.
Table 1.—Gear and apparatus used for the collection and processing of data on fish eggs and larvae in the California Current region.

<table>
<thead>
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<th>Gear or apparatus</th>
<th>Figure</th>
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<td>Specification in figure legend.</td>
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<td>Specifications in figure legend.</td>
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<td>Cod end</td>
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<td>Source of supply — Scripps Institution of Oceanography, Research Support Shop. Available as illustrated or with 2 bolts and eyes. When ordering specify cable size for which grooves will be drilled by supplier.</td>
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<td>Specification in figure legend.</td>
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<td>Available from scientific supply houses.</td>
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<tr>
<td>Measuring rule for fish larvae</td>
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<td>Board for calibrating flowmeters</td>
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with drum capacities of at least 30,000 ft (9,000 m), 3/16-in (5 mm) steel wire with a strain capacity of approximately 22,000 lb. (10,000 kg) — Figure 2. Some winches may be capable of handling conductor cable for telemetering data from all depths to which gear may be lowered. The NMFS research vessel, David Starr Jordan, is so equipped, with its standard
hydrographic winch starboard and its conductor-cable winch aport. Additional winches for trawling operations or very heavy gear are located below decks with their wire coming above decks to be fairleded astern to a powered H-frame and a stern ramp. Although the hydrographic-net tow wire is capable of the strain noted above, the boom supporting it (Figs. 2 and 3) can and need only support 5,000 lb, (2,200 kg). The block (Fig. 3) through which the wire is run also is capable of supporting 5,000 lb, and has three wheels of which one is a tenziometer.

To enable handling gear clear of the side of the ship, an overside platform (the “bucket”) — Figures 4 and 5—is another essential aid to data collection. It may be designed to be swung laterally outboard as on the Jordan or swung upward and outboard onto a ship’s rail to be fastened and kept there while at sea. The bucket is constructed of heavy steel, and its bottom is usually a steel grill which can drain immediately if water is shipped into it. A 100-lb, weight (45 kg)—Figure 4—is always attached to the end of the cable to aid in lowering the plankton net and the hydrographic cast of Nansen bottles.

Laboratory space on a research vessel may be of varying degrees of sophistication in size and equipment. On a small ship, one space may serve for all functions but must be of sufficient size to accommodate a number of Nansen bottles with thermometers and space to read them, an area with running salt and fresh water and sink for preserving biological specimens and an area for preserving water samples and determining their chemical constituents.

A survey for plankton sampling, extending to 14 or more days, will yield large numbers of biological samples for processing ashore. Adequate storage facilities must be available to keep them until off-loaded.

The Plankton Tow

The objective of the plankton tow is to obtain qualitative and quantitative samples of the zooplankton to the depth sampled at the time and place of the tow. Most important to the objective are the proper readings and recordings of the flowmeter, the recordings of the various parts of the towing times and wire angles, the necessity for the smooth paying out and retrieval of the net, rinsing the net, and the preservation and labelling of the sample.
Figure 3.—View of boom and block, upper left (also see Fig. 2), on R/V Jordan. Plankton net tow in progress. Telemetering inclinometer (see also Fig. 8 and 9), used to indicate wire angles, hangs from towing wire. Slack in wire indicates that the net is being lowered. Man on right is the winch operator at winch console, close to rail where he can watch net operations during launching and retrieval. Man on left will record wire angles when net is retrieved (see plankton tow procedure).
THE GEAR

*Plankton nets.*—The quantitative plankton collections of the CalCOFI are made chiefly with nets which retain organisms $\geq 500$ microns and $\geq 333$ microns.

*Note:* Most CalCOFI plankton tows were made with a 1-m silk or nylon net (ca. 0.5-mm. mesh). The directions for the tow indicate possible single or double rig.

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**Figure 4.**—Weight (100 lb. = 45 kg) being swung overside at end of towing cable. The weight is a cast-iron tapered cylinder 7½ inches (19.0 cm) in diameter, 13 inches (38 cm) high with its taper beginning about 9 inches (22.8 cm) from the bottom. One-inch (2.54 cm) stock is anchored in the casting and an eye, 1-5/8-inch (4.5 cm) I.D., of the same stock is welded to it. After casting, the unit is heavily galvanized. The diameter of the eye is enough to take a strong shackle and a heavy line through it. Note swivel between the shackles which keeps the weight from twisting the wire. Casing, partially shown at lower right, is used to store the weight when not in use. Overside platform at left (also see Fig. 5 and 12) shows grill floor (description in text).

**Figure 5.**—Overside platform (“bucket”) described in text, (also see Fig. 4 and 12), used to facilitate handling gear over water and away from side of ship. (The net is the Soutar-Hemingway Animal Trap (SHAT) used for vertical tows; it is not used or described here for standard collections for CalCOFI egg and larva data.)
Smith' described the plankton nets used on the surveys in this region and their history of design and material changes from 1939 to 1969. The standard CalCOFI nets presently used have \( \frac{1}{2} \)- and 1-m mouth openings and are made of nylon mesh (see Table 1 for source of net specifications and Fig. 6 for description of mesh sizes and assemblies for making plankton tows with either or both nets). The detachable cod end, also of nylon mesh, in which plankton is concentrated during a tow is illustrated and described in Figure 7 (source of specifications, Table 1). The adapter to couple the cod end to the net is described in the caption of Figure 7.

Smith, P. E. Plankton sampling nets and their data on surveys off California and Baja California since 1939. (Unpublished manuscript.)

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Figure 6.—Standard CalCOFI nets, 1-m mouth opening, 505-\( \mu \) mesh throughout, and 0.5-m mouth opening, 333-\( \mu \) mesh throughout (see Table 1 for source of specifications). Detachable cod ends are described in Figure 7. Flowmeters (see Table 1 for source of supply) are suspended in the mouth opening (also see Figures 13 and 15) by cloth-covered rubber shock cord. The cord is permanently attached to small metal eyes welded to the mouth rings.

Attachment to the lugs of the meters are by means of brass snap hooks. The net rings and braces between them are 1-inch I.D. (25.4 mm) galvanized pipe. Large ring is 1 m I.D., small ring is 0.5 m I.D. Nets are tied to the rings with 1/8-inch (4 mm) cotton cord. The three-way bridle is made up of 1/2-inch (12.7 mm) line (hemp, nylon, or diamond-braided polyethylene). Apex of bridle is a 2- or 3-inch I.D. (51 or 76 mm) steel ring of 3/8-inch (9.5 mm) thickness. The apex is 1 m away from the frame when the lines are stretched evenly. (Note: The points of attachment of the bridle to the rings must be no farther apart than shown. If greater than shown, the strain during any tow might bend the pipes to which the rings are welded.) The lead line, usually nylon or hemp, is 3/4- to 1-inch (20-25.5 mm) diameter for easy grasping when pulling nets aboard. (Note: A swivel should be attached between the lead line and the apex of the bridle and another swivel located at the cable clamp—see Fig. 12).

If either net is used alone (see Fig. 14) as has been the case in many years of CalCOFI tows with the 1-m net, the same size rings with the same specifications are used with a three-line bridle attached at equal distances on a ring. For the 0.5-m net a lighter lead line may be used, e.g., 1/2 inch (12.7 mm).
Figure 7.—Cod end (left)—(see Table 1 for source of specifications) used to concentrate plankton collected by net, detached from plankton net (right). Both are fastened with hose clamps to an adapter made of polyvinyl chloride (PVC) pipe. The pipe is approximately 4 inches (127 mm) long, 3-1/2-inches (95 mm) O.D. with 1/4-inch (6.5 mm)-thick wall. It is turned down about 3/32 inch (2.5 mm) along its length leaving flanges at each end to keep hose clamps from slipping off during tow; dimensions are not critical when turning the pipe.

**Flowmeters.**—When the data on plankton are processed, plankton volumes are calculated as ml/1,000 m$^3$ of water strained (see section on standardizing data). Water strained through the net is measured by a flowmeter suspended in the mouth of the net (Figs. 6 and 13)—see Table 1 for type of meter used and source of supply. The meters are calibrated according to the methods described below in the section on standardizing data.

**Inclinometers.**—The CalCOFI standard plankton tow is taken by lowering the net(s) to the desired depth and retrieving it at a given rate while the ship maintains a speed that keeps the tow wire at an angle of 45°. The instrument that measures the angle of stray, the “wire angle,” is a quadrant called the inclinometer (Fig. 8) which, when hung on the wire (Fig. 3), can be monitored to record the wire angles and to regulate the ship’s speed to keep the desired angle.

Two kinds of inclinometers have been used. The one shown in Figure 8 (see Table 1 for source of specifications) is equipped with a telemetering device developed by Charles Forester, Master of the research vessel *Jordan*. When equipped for telemetering, the angle of stray can be controlled from the bridge or engine room where wire angles are indicated in microamperes (Fig. 9). Wire angles on the quadrant (Fig. 8) are recorded on the tow sheet by an observer on deck. The second kind of inclinometer is simply one without the telemetering device. In this case, an officer of the watch or the recorder observes the wire angles during the tows and signals the bridge or engine room if the desired angle is not being maintained.

**PROCEDURE FOR PLANKTON TOW**

*The data sheet.*—When the gear is assembled as shown in Figure 6 and the data sheet for the plankton tow is prepared, the tow is ready to
begin. The data sheet used here for the plankton tow (Fig. 10, specifications in figure legend) is preprinted to record data for tows up to 300 m of wire out. Twenty items are numbered for automatic data processing.

The tow.—Before the net-tow station is occupied, the following numbered items should have been recorded on the data sheet.

1-Cruise, 3-Date, 4-Order occupied, 5-Station, 11-Net number (regular and/or fine) this is usually the mesh-size number (see Fig. 10), 12-Meter number (regular and/or fine), 14-Carry-over, initial meter reading (regular and/or fine).

The net tow is made off either side of the ship as follows: (Tows off the stern are not recommended because of turbulence from ships' screws.)

1. The ship is stopped; the station depth is requested from the bridge and recorded in the lower left hand section of the data sheet.

Note: With 300-m wire out, and the wire angle at 45°, the net is approximately 210 m deep. (Wire out X cosine of 45°—0.707—= net depth.) If station depth is less than 130 fathoms (238 m), a “Depth-of-Tow” graph (Fig. 11) is referred to in order to determine the proper amount of wire to pay out so that the net and gear will not hit the bottom. Shallow and deep tows are payed out and retrieved at the same rate as routine standard tows.

For shallow and deep tows, other than routine (items 20-22), the section 20'-22' should be used, recording lengths of wire out in decrements of 10, as called by the winch man, and wire angles (see below, procedure for tow, item 7a) at such calls.
2. The flowmeter is read and checked against the recorded initial meter reading—item 14. If there had been a previous tow, this should have been the final meter reading—item 13 on the previous tow sheet. If the reading changed between tows, the last recording is crossed out, the new reading entered, and an explanation given in the Remarks section, lower right hand part of the data sheet.

3. The 100-lb. weight is lowered about 10 to 15 m below the surface of the water. If the ship is still slightly underway, the wire is pulled to the side of the bucket and fastened close with snap hook attached to the outside of the bucket (Fig. 12).

4. The bridle clamp (Fig. 12)—see Table 1 for source of supply—is fastened tightly to towing wire, and a safety chain or line (Fig. 12) is fastened to the wire above the clamp. The clamp should be about 15 to 20 m above the weight, less in shallow water (see Depth-of-Tow graph—Fig. 11—for directions concerning weight in shallow tows).

5. The inclinometer is fastened to the wire above the clamp. Enough slack is left on the line to the inclinometer so that when the proper angle is achieved during tow (Fig. 3), it will not ride up on the cable to hit the block. (If the survey is for net tows only, the inclinometer may be left "permanently" on the tow wire. It is always removed if a hydrographic cast has to be made with the cable.)

6. The cable clamp is lowered to the sea surface, and the winch meter is zeroed.

7. The ship is set underway, wind off the bow on the side on which the tow is taken, and the signal to start the tow is given from the bridge. The block or pin, which keeps the blades from revolving between tows, is removed from the meter(s), and the net(s) is thrown into the water (Fig. 13). (Some
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<th>CRUISE</th>
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<th>DATE</th>
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### NO. OF ANGLES

- TOTAL: 3
- ACCEPTED POSITION
  - LATITUDE: 24°30' N
  - LONGITUDE: 120°30' W

### ROUTINE

- TIME NET ENTERS WATER: 02:24

### ANGLES WIRE OUT

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### OTHER

- TIME NET ENTERS WATER: 
- ANGLES WIRE OUT
- ANGLES WIRE OUT
- ANGLES WIRE OUT

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### INCHES OF PLANKTON

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### DEPTH

| 163  |
| FATHOMS |

**Figure 10.—Plankton-tow data sheet.** A sample copy of a sheet made out for station 80.52 on cruise 6907-J (see text). The sheet is a water-resistant linen designated "36-lb. ledger." Other qualities of paper, not so designated have been found to deteriorate when wetted by rain or handling. The sheets are delivered in pads glued at one end. When readied for use, two sheets at a time are torn off, a carbon paper inserted between them, and all are placed on a clipboard and held at the bottom of the sheets by a heavy rubber band.
meters have "automatic" blocks that release the impeller blades when water flows through them.)

a. The stop watch is started when the flow-meter (s) is seen to sink below the surface of the water.

Figure 11.—Depth-of-tow graph.—To be used if station depth is 130 fathoms or less when standard tow is 300 m wire out. If standard tow is less than 300 m wire out, e.g., 100 m or 200 m, the graph is still applicable. For 100-m standard tow, use graph when station depth is 45 fathoms (82 m) or less. For 200-m standard tow, use graph when station depth is 165 fathoms or less. Except for the very shallow depths, 20 fathoms or less, where a tow is practically all at the surface, the points on the diagonal line indicating the amounts of wire to be paid out are calculated to keep the net approximately 10 to 18 m off the bottom. This is considered a safe margin, and it is recommended that no changes be made when one considers that only the rocking of a ship in heavy seas can reduce an 8-m safety margin to 5 or 3 m.
b. The net(s) is allowed to stream out (Fig. 14) before lowering, and when it is obvious that it is not tangled, the wire is payed out at 50 m/min until the desired depth is reached.

c. At the desired depth, the watch is stopped, sinking time is recorded (item 8), and watch zeroed and restarted immediately.

Note: Since the net is "fishing" on the way down, sinking time is as important as that of retrieval. Recording the time is simplified if "Time Net Enters Water" is recorded to the nearest 5 min. (This is the item near 20—or 20’—Routine or Other, depending on tow—see later). This time is also recorded at item 6.

d. When the watch is restarted the net(s) is left at the desired depth for 30 sec (hypothesized that a "falling" net(s) will straighten out at depth in the 30-sec interval).

e. At the end of 30 sec the watch is not stopped, the angle is recorded for that depth, and retrieval is begun at the rate of 10 m per 30 sec. The angle is recorded at every 10 m in items 20-22 (Routine) or 20’-22’ (Other).

Note: Ship speed, during sinking, times at depth, and during retrieval, is maintained to keep the wire angle at 45°. In dead calm, it may be necessary to run the ship in circles to maintain the wire angle.
f. The net(s) is brought directly out of the water at a steady rate. When the flowmeter (s) breaks the surface the watch is stopped. Its reading is the towing time (item 9).

8. The net(s) is rinsed to get all the plankton into the cod end, keeping the cod end(s) dangling and the net ring(s) at rail height (Fig. 15). The net(s) is brought aboard (Fig. 6), and the cod end(s) removed, keeping the plankton sample(s) from spilling back into the net(s). The plankton is preserved immediately.

9. The plankton is poured from the cod end into a jar of appropriate size, usually a quart. The cod end is rinsed down to gather the last of the plankton at its bottom. When fairly well drained, the cod end is everted over and into the jar, and the remaining plankton washed off carefully.

10. The preservative (50 ml full-strength formaldehyde per quart) is added (see note below) when the jar is at least three-fourths full of seawater plus plankton in order to avoid “burning” the delicate planktonic organisms. Buffer to counteract the
11. Inside and outside labels are filled out. (The greater part of these might be filled out before a station is occupied.) Inside and outside labels are illustrated in Figure 18. If more than one jar is used for a sample, labels are so designated—1 of 2, 2 of 2, or 1 of 3, 2 of 3, etc. The number of jars used

is noted in the proper space on the tow sheet (see 14d below).

12. The cod end(s) is washed (it may be left everted) and replaced on the net(s) in preparation for the next tow (Fig. 6).

13. Before leaving station the meter(s) is read and recorded as the final reading at item 13, and the initial reading, item 14, is subtracted from item 13.

acidity of plankton in Formalin is added—20 ml per quart—(saturated solution of sodium borate in seawater). The jar is filled almost to the top with seawater, capped, and shaken to insure a good mixture of preservative and plankton.

Note: Full-strength formaldehyde aboard ship is kept in 5-gal polypropylene carboys (see Table 1 for source of supply). With the carboy moored securely above the sink (Fig. 16), the preservative is drawn by siphon action. A further safety measure now adopted is to draw the formaldehyde via a teflon tube into a 50-ml plastic syringe through an automatic double valve (Fig. 17)—see Table 1 for source of supply. The buffer is added with a 20-ml plastic syringe fitted with cannula (a "needle" without a point).

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Note: Experience will teach the readers what a normal meter reading should be for a standard tow. If meter readings are not normal, the net tow may have to be repeated. A very high reading may have been caused by too great a ship's speed—check for many high wire angles. A low reading may have been due to too slow a ship's speed—check for many low wire angles. Another reason for low meter readings may be clogging of the nets. This may be cumulative if a net is not rinsed properly or it may occur at a single station. If a meter shows a trend toward lower and lower readings, it is not malfunctioning, and the net should be washed (see below). The net tow need not be repeated if it is obvious that heavy clogging is the reason for low readings (it will only clog again) or if the ship's speed has caused low or high angles. If wire angles are normal, the net clean, the towing time routine but the meter reading is low, the cause

could be that a bit of detritus, a fish, or even a large jelly or salp had become entangled in the meter blades for a portion of the tow. Under these conditions, the tow should be repeated. If the reading is again very low and it is obvious that the flow-meter is not functioning properly, replace the meter and repeat the tow. Do not oil or grease any meter or make any repairs that might alter the rotation of the blades. Repairs of this type would seriously affect the calibration of the meter.

14. The tow sheet is completed as follows:

a. Towing time—the time at which the watch was stopped when the meter(s) broke water, is recorded at item 9.

b. Total towing time—recorded in item 10, is the sum of items 8 and 9. In a 300-m tow, total time should be about 21’30” —6’ sinking time + 15’30” towing time. If total time is off by 15 to 20 sec, it must be explained in the Remarks section. The most usual variation will be in the sinking time, caused by a slightly faster or slower rate in paying out the wire than the recommended 50 m/min. In certain conditions, such as poor control of the ship, countercurrents below the sea surface adversely controlling the net as it falls, etc., the winch operator may have to depart from the sinking-time procedures to slow the falling net in order to keep it from becoming tangled. Such departures from normal procedures must be recorded in the Remarks section.

c. Total towing time is added, in minutes and seconds, to item 6 to record the hour, minutes and seconds at item 7. This is actually the time the net comes out of the water.

d. Number of jars per sample—lower left hand section of sheet.

e. Inches of plankton—this gives approximate volume before water is added.

f. Formalin and borate added—the person who adds the preservative and buffer should initial this box for each net—after the Formalin and borate are added.

g. Sample labelled—the person who labels the sample(s) should initial this box —after the sample(s) is labelled.

h. Depth, wind, sky, sea swell—should be given by a crew member to the observer who is recording the angles while the tow is being taken.
Figure 17.—Two-way, double action valve on syringe (see Table 1 for source of supply). Usually the valve is fixed to the syringe by a screw-on action. In this illustration, the valve is fitted tightly by friction to a disposable plastic syringe.

Figure 18.—Labels for plankton samples. Clockwise from upper left—Tie-on cardboard label on 1-quart (1 liter) bail-type jar; screw-on cap labeled with Martek pen, on 1-pint (1/2 liter) jar; self-adhesive label on screw-on cap; inside label on which information duplicates (not in any of these cases) whatever is written on an outside label. Inside label is made of 32-lb., chemically resistant linen called “Resist-all.”

i. Amount of clogging—should be checked in one of the appropriate boxes. This is best observed by noting the variation of the meter readings (see 3b above). If washing is needed, one of three methods may be used: (1) The net is everted, still on its ring, and brushed down with an ordinary sweeping broom and running seawater; (2) rings are stood on edge, net is tightened along its length by tying down end (without cod end attached) and hosed down with high-pressure fire hose. This is very effective provided that plankton has not dried in the meshes; (3) net is detached from ring and put in a washing machine using a 30-min cycle, warm water (not hot) and a nonpolluting detergent.

j. Rips and holes in the net—the net should be looked at after every tow to check on needs for repair or replacement. If holes or tears are small, they should be sewn before the next tow with nylon thread of a dark color (to be easily located for sewing machine repair later on shore). Check appropriate boxes. If
the net is torn beyond mending at sea, replace the net.

k. Recheck sheet to be sure that all items are filled in. Nonroutine items should be included in Remarks section, e.g., odd meter readings, prolonged stops at stations, delays between stations, etc.

l. The accepted position, item 16, may be listed when the station is occupied or at the end of the cruise when the captain has compiled a complete list of the positions of all stations.

m. The occupancy code, item 2, is filled in, onshore, at the end of the cruise. This is usually one of a series of numbers used by the programmer to describe the type of tow or the station occupied.

15. The sheet is set up for the next station: items 1, 3, 4, 5, 11, 12, and 14 are entered. Item 14 should be the final reading of the preceding tow and should be rechecked before starting next tow.

### Additional Data Collections

A tow for plankton is made at every station occupied during a CalCOFI survey. Additional work and data collection at each station include: Collection of meteorological data; a bathythermograph with the expendable bathythermograph (XBT), a surface temperature reading taken with a bucket thermometer, a sample of water from 10-m depth with a Nansen bottle from which shipboard analysis is made of salinity and nutrients including phosphate, nitrate, nitrite and sulphite; drift bottle releases at specified station, secchi disc reading at all day stations and Pacific saury (*Cololabis saira*) observations at all night stations.

The XBT is dropped from a launching tube, located aft near a ship’s rail, to record temperature profiles to 1,000 ft (450 m) deep. The recording is made electronically in a sheltered space (laboratory, etc.) on the vessel (Saur and Stewart, 1967; Saur and Stevens, 1972). The XBT has replaced the former BT which was launched and retrieved on a cable with a small winch located aft near a ship’s rail recording temperature profiles on a smoked slide to 450 or 900 ft (137 or 355 m).

### PROCESSING PLANKTON AND STANDARDIZING DATA

Processing of plankton is begun at the laboratory when the collections are brought back from sea. This is carried out in several steps: Measuring the volume of plankton in each sample, sorting out and enumerating all fish eggs and larvae, identifying all larvae, measuring certain larvae, identifying certain fish eggs and staging (ageing) some, and finally, curating all fish eggs and larvae. All data are standardized (see below) and now are subjected to automatic data processing (ADP) for final analyses and publication. Two methods for standardization are described, one, the old method of hand calculation and two, the ADP which are described wherever changes have been affected, and for which a flow diagram is depicted in Figure 19.

**Plankton Volume Determination**

Plankton volumes are determined by displacement, (sometimes termed “wet volumes”) recorded to the nearest milliliter. Two volumes are recorded for each sample:

1. **Total volume**—includes everything in the sample except small adult fishes, juvenile fishes, squid, octupi, and adult pelagic crabs (*Pleuroncodes*) none of which are considered planktonic.

2. **Total volume minus large organisms**—large planktonic organisms are jellies and tunicates whose individual volumes exceed 5 ml.

The plankton volumes for a cruise are recorded on a Plankton Volumes data sheets (Fig. 20) beginning with data from the original tow sheets, first listing all the stations occupied in their numerical order and noting the number of jars used for each sample. The plankton samples are removed from their boxes and readied for measuring their volumes by arranging them in the numerical order of stations. The procedure for determining volumes is as follows:

1. Each quart jar sent to sea is calibrated and etched with a number (see Fig. 22) that represents its total volume when filled to a level at which a mark on one device (a float) matches a mark on another (a T-guide)—Figs. 21 and 22. When the calibrated jar contains a plankton sample, the
Figure 19.—Flow diagram of procedures by the National Marine Fisheries Service for the processing of plankton, fish eggs and larvae, and data through tape storage, La Jolla, Calif. (Designed by J. R. Zweifel, NMFS, La Jolla). See text for disposal of sorted samples.
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</tbody>
</table>

Figure 20.—Plankton-volume data sheet.—A sample copy of a sheet made out for stations 60.50 to 70.80 on cruise 7003-"Ogon." (This does not match any of the sets of data for which examples are shown in this report.) The data shown here were collected on a special cruise on the CalCOFI grid by a cooperating vessel, Ogon, of the U.S.S.R., to show fractioning of samples (no longer done at this laboratory—see note on description of Set IV in the section on processing data.) These samples were fractioned by agreement with the Russian participants in that each organization would keep one half of each sample for study at its respective laboratory and exchange the data on all samples.

The first step in measuring is to bring the plankton plus its preservative to the “proper level” by using the T-guide and float as shown with Figure 22. (Alternative method—If calibrated jars, T-guide, and float are not available, the sample and its preservative are poured into a graduated cylinder and enough preservative is added or subtracted to bring the level of the liquid to an even milliliter, usually 900 or 1,000; the volume is recorded.)

2. A funnel is placed in a clean graduated cylinder (see Fig. 24 for specifications), and a 333-μ-mesh nylon draining cone (source of specifications—Table 1) is placed in the funnel.

3. The plankton and preservative in Step 1 (either alternative) are poured into the draining cone (Fig. 23). The plankton is retained in the cone while the liquid drains into the cylinder. The plankton is considered drained when the liquid from the bottom of the cone diminishes to an occasional drop. Draining time varies with the size and composition of the sample.

4. The volume of the drained liquid in the cylinder (Fig. 24) is subtracted from the initial volume of plankton plus liquid (Fig. 22)—calibrated quart jar or graduate in alternate of step 1. The difference, the total volume of the sample, is recorded on the data sheet.
Figure 21.—T-Guide and float, actual size (designed by J. R. Thrailkill, NMFS). This is used to calibrate quart jars and to adjust volumes of plankton + preservative in measuring plankton volumes. To use: the float, triangular section pointing up, is inserted between the vertical limbs of the T-guide. The apparatus is placed in a calibrated jar of plankton plus preservative and the horizontal limbs rested on top of the jar (also see Fig. 20). Liquid is added or subtracted causing the float to move up or down until the etched line on the float is between the etched double line on the T (see Fig. 22). The coincidence of the lines indicates that the proper level has been reached, i.e., the volume, in millimeters, etched on the jar—accuracy ± 1 ml.

5. Large jellies and tunicates are removed (estimated as equal to or larger than 5 ml in volume), washed, and placed into a graduate with a known volume of 5% buffered Formalin. The difference in this reading and the known volume of added solution is the Volume of the Large Organisms which is subtracted from the Total Volume and recorded as Total Volume Minus Large Organisms (Fig. 20).

6. The drained plankton and the large organisms are put into a pint jar(s)—see Note below, and the jar(s) is filled with the original preservative. The original inside label(s) is placed in the jar(s). If a screw-top quart jar is used at the time of collection, the same lid is used on the pint jar. If that cap was marked with a writing pen (Martek) or a self-adhesive label, it is left as is. If a string label was used, as on a bail-type jar, the top of the cap is labelled with a self-adhesive label or Martek pen (Fig. 18).

Note: Pint jars are used to store measured plankton because it has been found that almost 90% of all single-jar samples on CalCOFI surveys will fit into pint jars. Even if more than one pint jar is used for some samples, the boxes for storing this size jar require very little more than half the storage space needed for quarts.

7. Calibrated quart jars - with unmarked lids - are returned to wooden seagoing boxes (ca-
Figure 22.—T-guide in use in first step in measuring plankton volume. The jar is calibrated to 892 ml, liquid alone or liquid plus plankton. Liquid is being removed to match the etched lines on the float and T-guide (see Fig. 19 for directions for use).

Capacity: 12 jars in eggcrate partitions) constructed of 1/2-inch (12.7 mm) plywood with hinged lids. If laboratory and storage space on shipboard are reasonably dry, the original cardboard containers can be used to store jars of plankton at sea.

8. Pint jars with plankton are packed in numerical order of stations into cardboard cartons, and the outside of the cartons are labelled with cruise number and numerical listing of the stations in the box stored to be visible on shelves.

**Plankton Sorting**

Each measured sample delivered to the sorting laboratory is sorted for all fish eggs and larvae of which all are enumerated. Some fish eggs and larvae are identified, some are scanned and categorized as “few”, “many”, or “abundant”, and some fish larvae are measured (Table 2).

Although techniques may vary with individual sorters, the general method for sorting is as follows:

1. The plankton is removed from its preservative by straining it through a 333-μ mesh nylon draining cone (the same kind used when measuring plankton), and the plankton, with about 2 liters of fresh water plus a few drops of full strength formaldehyde, is put into a 3-liter beaker. The original preservative is kept in its original jar. Fresh water is used because it has been found that prolonged exposure to concentrations of Formalin in handling, stirring and under their eyes, even 3 to 5%, may cause sorters to become sensitive and allergic to fumes and liquid. A sample can be kept as long as 1 month in the weak solution of fresh water with formaldehyde. This does not imply that such length of time is necessary for sorting any single sample. The average sorting time is about one sample (100 ml plankton) per day per sorter.

2. The plankton is stirred and poured into a small beaker (200 ml). This, in turn, is stirred and poured into a number of Syracuse dishes (ground-glass sides) from
Figure 23.—Pouring plankton and preservative into draining cone is the third step of measuring plankton volume. The cone (333-μ nylon mesh—see Table 1 for source of specifications) retains the plankton; the cylinder receives the fluid. Large sewing needles suspend cone in funnel keeping mesh from touching funnel’s sides and allowing proper drainage.

which the organisms will be sorted. These dishes are aligned on one side of the microscope. On the other side of the microscope are a number of Syracuse dishes, each labelled on its ground-glass surface with the name of the organism which will be transferred to it when sorted (Fig. 25). Each of the labelled dishes is about half full of 3 to 5% buffered Formalin. (In this instance, the few dishes of such Formalin are not enough to affect the sorters adversely.)

3. Using a binocular, dissecting microscope, usually at a total of 9X power, with trans-

Figure 24.—Reading volume is the fourth step of measuring plankton preservative after plankton has drained to an occasional drop. The volume of the calibrated jar, holding plankton and preservative (Fig. 20), less the volume in the cylinder equals the displacement volume of the plankton. The cylinder is plastic tubing, 1-1/2 inches I.D. × 34 inches long (3.8 × 86.4 cm) with graduations etched on the cylinder or on a grooved board, as illustrated. The graduations on cylinder or on the board are 5-ml units from 0-600 ml and 2-ml units from 600-910 ml. Volumes are read to the nearest millimeter.
Figure 25.—Arrangement of plankton sorter's work. Unsorted plankton on left side of microscope, sorted organisms on the right. (See text for procedures.)

mitted light, all fish eggs and larvae are picked out with pipettes and fine quality (stainless steel) forceps and transferred to their appropriately labelled dishes.

Note: Research quality microscopes are used because any of inferior quality would be detrimental to the eyesight of persons engaged in this type of week-long work for 6 to 8 hr per day.

4. When the fish eggs and larvae are sorted from a dish, its remaining contents are poured into a 3-liter beaker labelled "Sorted."

5. Each dish of sorted organisms is checked for final identification when its contents are enumerated and/or measured as noted in Table 2. Fish larvae are measured to the nearest one-half millimeter on a transparent millimeter rule (specifications below). Each measured species is tabulated on the form illustrated and described in Figure 26.

Note: The scale for measuring fish larvae is a transparent plastic rule, about 0.5 mm thick, on which the markings, in millimeters, are etched on the plastic. A piece of the rule, about 50 mm, is taped between two thin pieces of glass—usually standard microscope slides 76 × 51 × 1 mm. The larvae are piled in a small mass on the slide, and individual specimens are gently dragged over the top of the scale with a clean dissecting needle. After measuring, they are dragged away into another pile until all are measured. Finally they are placed in a vial and labelled.

6. The sorted plankton (invertebrates) is poured into the mesh cone to drain off the water, and the plankton is returned to its original jar and preservative. (When a cruise is completed the samples are sent to SIO for curation and study of selected invertebrates.)

7. When the checking in step 5 is done another form, the sorter's work sheet (Fig. 27), is filled out listing the numbers of organisms of each type sorted. The form also includes other information as illustrated. Whole larvae, head sections and tail sections are listed and totaled for all species measured and for other fish larvae. Totals listed for measured larvae should be the same as those on the tabulation sheets; (exceptions noted in caption for sorter's work sheet—Fig. 27) the totals of head and tail sections should equal the DIS on the tabulation sheets (Fig. 26).
Table 2.—Organisms identified and enumerated and/or measured during sorting of CalCOFI plankton samples.

<table>
<thead>
<tr>
<th>Species</th>
<th>Common name</th>
<th>Labelled</th>
<th>Enumerated</th>
<th>Measured</th>
<th>Scanned</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sardinops caeruleus</td>
<td>Pacific sardine eggs Larvae</td>
<td>Sardine E</td>
<td>X</td>
<td>X</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td></td>
<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Engraulis mordax</td>
<td>Northern anchovy eggs Larvae</td>
<td>Anchovy E</td>
<td>X</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td></td>
<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Merluccius productus</td>
<td>Pacific hake eggs Larvae</td>
<td>Hake E</td>
<td>--</td>
<td>--</td>
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<tr>
<td></td>
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<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cololabis saira</td>
<td>Pacific saury eggs Larvae</td>
<td>Saury E</td>
<td>X</td>
<td>--</td>
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</tr>
<tr>
<td></td>
<td></td>
<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Trachurus symmetricus</td>
<td>Jack mackerel eggs Larvae</td>
<td>Jack mackerel E</td>
<td>--</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td></td>
<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Etrumeus acuminatus</td>
<td>Round herring eggs Larvae</td>
<td>Etrumeus E</td>
<td>X</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td></td>
<td>L</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other fish eggs</td>
<td></td>
<td>OFE</td>
<td>X</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td>Larvae</td>
<td></td>
<td>OFL</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

1 To nearest one-half millimeter.
2 Categorized.
3 Occasionally.

8. Each group of organisms is placed in a 2-dr vial with screw cap. An appropriately marked label is placed in each vial, and the vial is capped. Each label includes cruise number, station number, date of tow, organism name and total. If organisms were measured this is indicated on the top of the cap on a self-adhesive label. Labels are written with pencil (grade equal to “H”) or waterproof ink (Higgins Engrossing Ink, No. 892, which does not clog the type of pen used here—Kohinoor Rapidograph, No. 0 or 00).

Note: Paper for labels should be 100% rag content. Inferior quality paper eventually deteriorates in Formalin or loses legibility.

The type of screw cap used here is plastic with a vinyl insert (Fig. 28) that is “self-sealing” when screwed tightly on the vial, thus preventing evaporation for long periods of time and decreasing amount of curating time needed to replenish evaporated preservative (see section on Curating). This is much preferred over corks, rubber stoppers, or screw caps with paper liners.

9. All material is checked including the work sheets and vials with labels.

10. A sorter’s master sheet (Fig. 29) is compiled on which the sorting data are summarized for each station from each sorter’s work sheet.

11. The vials of each station are banded and arranged in numerical order of stations in small cardboard boxes for delivery to the identification group and further work.

Identification of Fish Eggs and Larvae

When the identifiers receive the vials and master sheets from the sorting laboratory their procedures are generally as follows:

1. All eggs and larvae identified and enumerated by the sorters are verified, and their numbers listed in the appropriate boxes at the tops of the columns on the form illustrated in Figure 30. Since our primary interest is in certain commercial species, those are prelisted as shown on the form.

2. The OFE’s (see Table 2) are verified. If any of the form’s prelisted species have been overlooked by the sorters, they are listed and added to the totals on the form shown in Figure 30. (Also see Figure 32 for method used for automatic data processing.)

3. The OFL’s (see Table 2) are identified according to the classifications of Berg and listed in the large space on the form. Sebastodes = Sebastes (rockfishes)—not the responsibility of the sorters—if present, are totaled and listed in the appropriate box. Flatfishes, some of which are com-
mmercial species, if present, are listed first on the form, then all other OFL's and myctophids. The methods of identification in the taxonomy of fish larvae, worked out by E. H. Ahlstrom of our laboratory, are too involved to be discussed here. Sufficient to say that the larvae of at least 100 families are found in the California Current area. At present, approximately 100 kinds can be identified to species, 40 to genus and the rest to family. Size is an important factor in identification since identifying characters of larvae change with growth. The identifiers can recognize more than 1,000 sizes, shapes, pigment patterns, eye forms, etc., which characterize individual larvae of the various species.

During identification the OFL's are checked for overlooked species that are the responsibility of the sorters. In most cases of omission, the larvae are in such poor state that it may be impossible for the sorters to have identified them or so poor a state that they could not be identified at all. If identifiable, they are added to the other totals, where appropriate; if not they are put into a disintegrated (DIS) category. All identified fish are returned to one vial. Some, if in excellent condition, may be kept out for further study or the reference file.

![Table of larval lengths](image)

Figure 26.—Plankton sorter's tabulation sheets for measuring fish larvae—sample copy of one of two sheets used to measure anchovy larvae for station 80.52, cruise 6801, regular mesh (505 μ). The tabulations are made up on two sheets to include lengths of larvae by 0.5-mm sizes from 2.0 to 8.0 and 8.5 to 15.0 m with additional lines for longer larvae. At the bottom of the second sheet is a line for tabulating disintegrated larvae (DIS).
4. If sardine eggs are in the sample they are staged according to the method devised by Ahlstrom (1950) as to their state of development in 11 phases from fertilization to full development before hatching. Then they are aged according to the water temperature and time of day at which they were collected.

5. All labels are the same as those for plankton sorting. If larvae are bottled separately,
from the rest of the sample, this fact is noted on the enumeration sheet and on the back of the label in the vial in which they would have normally been placed. Vials are stored by numerical order of cruise and cruise station in specially constructed cardboard boxes $3 \times 5 \times 7\frac{1}{2}$ inches with lids, each containing 60 vials labelled appropriately on the outside of the box by category and cruise. Each cruise may have from 10 to 28 boxes depending on the abundance of organisms and time of year of a survey.

The boxes are stored by cruise in a special area in the laboratory.

REFERENCE COLLECTION

A special reference collection, the "larva library", is kept adjacent to the identification laboratory. The specimens in this collection are usually the "best of a kind" and used for reference in identifying questionable material or rare specimens whose characteristics may have been forgotten over long periods of time. The material is also used occasionally to train identifiers.
CURATING

The general and reference collections are periodically checked to assess the evaporation of preservative from the vials. The screw-top lid with the vinyl liner (Fig. 28, also see section on Plankton Sorting) now used on our vials is virtually evaporation-proof if well tightened when stored. Occasionally some vials with loose caps may lose liquid by evaporation in which case preservative is added, the cap tightened properly, and the vial stored again.

Standardization

During tows for plankton, different volumes of water are strained through the net depending on different speeds of the tow (more water with high speed) or different times of tow (in shallow versus standard depths).

In order to make tows comparable, all hauls are adjusted to a standard amount of water strained per unit of depth fished—10 m$^3$ of water strained per meter of depth fished. This value is used because it gives a factor of approximately...
1.0 for a net with a 1.0-m-diameter mouth opening. A standard haul factor (standardization factor) is derived for each haul by the following formulation:

\[ S = \frac{10D}{V} \text{ or } \frac{10D}{R \cdot a \cdot p} \]

where \( S \) = standardized haul factor (= SHF),
\( D \) = average depth of haul—derived by multiplying the cosine of the average angle of stray by the length, in meters, of the towing cable. (The cosine of the average angle of spray, derived from a tangent of all angles, is considered more representative of the haul as a whole than the cosine of the angle of stray at the greatest depth.)

\( V \) = total volume of water strained in cubic meters.

\( R \) = total number of revolutions of the current meter during the tow.

\( a \) = cross section area of mouth of net in m.²

\( p \) = length of the column of water, in meters, needed to effect one revolution of the current meter at average speed at which the haul was taken (determined from the appropriate calibration graph—see calibration of flowmeters).
CALIBRATION OF FLOWMETERS

For calibration, a meter(s) is hauled or pushed at different speeds over a measured distance. (Our personnel have developed a method for calibrating three meters at a time, fastening them to a bracket under a board—Figs. 33 and 34—and pushing them over the measured distance — 42 ft = 12.2 m.) Performance tests are made before (Fig. 35) and after every cruise, and a graph is constructed in which the independent variable is the length of a column of water needed to affect one revolution of the meter (meters per revolution) at any given towing speed. The graph (Fig. 36) applicable to a given cruise is based on the average of two calibration trials. The graph is curvilinear with the highest values of m/rev associated with the lowest numbers of rev/sec because of friction at low speeds. The curve flattens out with increase in speed and a corresponding decrease in friction.

With preparation by automatic data processing (ADP), the meter calibrations are key punched and programmed to produce a regression line where meters per revolution are plotted against seconds per revolution in order to linearize the relationship over the range of values obtained in all the plankton tows for which the meter was used.

Standardization of data for every station on a cruise is begun with the preparation of four “sets” (Fig. 37) as follows:

Set I — to derive the SHF from the formula

\[ S = \frac{10 \, D}{V} = \frac{10 \, D}{R \, a \, p} \]
Figure 33.—Board used to calibrate T.S-Flowmeters (one meter attached). The board is balsa wood, covered with fiber glass to make it waterproof, 4 ft long, 15 inches wide X 2-1/2 inches thick (151.9 X 38 X 6.4 cm).

Set II — to determine the volume of plankton collected per 1,000 m³ of water strained.
Set III — the station data and plankton volumes for publication.
Set IV — to adjust SHF to percent of sample sorted—no longer done—see note following description of Set IV below.

In addition, if an improper meter reading was obtained or was missing for any tow, a scatter diagram and regression line (Fig. 38) had to be plotted and calculated for all meter readings (ordinate) against all average tangents (abscissas) in order to obtain an estimate of a meter “reading” to apply to that tow (see parenthetical data for station 60.70 in Figure 37).

Each set duplicated the cruise number, the ship, the dates of the cruise—start to end, meter number(s) and net number(s).

On Set I, 14 columns are used of which the last is the SHF. The column headings are as follows:

1. Station number—from tow data sheet.
2. Order occupied—from tow data sheet.
3. Total time of tow in minutes and seconds—from tow data sheet.
4. Total time in seconds—derived from 3.
5. Current meter revolutions, difference in final and initial reading—from tow data sheet.
6. Revolutions per second current meter, from 5, divided by time in seconds.

Figure 34.—Brass frame under calibration board (see Fig. 33) on which to hang flowmeters. Only three meters are calibrated at one time, the hangers nearest the board are not used; meter readings were found to be erratic because of the turbulence in that area. The brass frame is 8 inches X 14 inches (20.3 X 35.5 cm) made of four pieces of brass welded at their ends. The brass material is 3/16 inch X 2 inches (5 X 51 mm). The brass angles on which to hang the meters are cut from 1/8-inch (3 mm) stock, 1-1/4 inches (32 mm) each side, 3 inches (72 mm) long. Holes are bored to conform to distance on centers of holes in meter lugs. Nuts are welded in position to take bolts that slip fit through meter lugs. Meters just hang on the bolts.

7. Average tangent—derived from tangents of all angles taken during tow.
8. Calibration factor = meters per revolution, the p in formula above—taken from calibration curve (Fig. 36).
9. Calibration factor X areal cross section of net = a constant which, when multiplied by R (total revolutions/haul) gives
10. Volume of water strained = Col 5 X Col 9
11. Cosine of average tangent from Col 7
12. Wire out = total wire out before net is hauled in
13. STD haul factor = \( \frac{\text{Col 13} \times \text{Col 10}}{\text{Col 10}} \)
Figure 35.—Calibration of flowmeter—sample copy of record for meter #1179 on June 21, 1966. The meter, with others on a board (see Fig. 33 and 34), is pushed over a measured distance at a number of speeds, in this instance 13 runs at approximately 15 to 40 sec. The meters per revolution are plotted against revolutions per second for part of the calibration curve shown in Figure 36.
Set II derives the plankton volume per 1,000 m³ water strained by the following formula:

\[ V_{p/m^3} = \frac{V_p}{V_w} \times 1,000 \]

where \( V_{p/m^3} \) = plankton volume per 1,000 m³ water strained.
\( V_p \) = plankton volume collected in net tow (from plankton measured).
\( V_w \) = cubic meters of water strained (from Set I).

Set III combines all the station data for publication.

Set IV adjusting for percent plankton sorted multiplies the SHF by reciprocal of the fraction sorted, e.g., 50% (one-half) of sample, multiply SHF by 2, 25% (one-fourth) sorted, multiply SHF by 4, etc.

Note: It has often been found that when some organism such as rare and/or larger fish larvae are scarce, fractioning a sample tends to give an erroneous count of actual numbers present, where it is highly probable that the few specimens in a sample can be left in the unsorted portion of the plankton. Therefore, it has been our policy of late to sort 100% of all samples.

With automatic data processing only two sets are printed out, one to obtain volumes of water strained, the standard haul factor for each plankton haul as for Set I above, and the other to produce station data and plankton volumes as for Set III above.

These data sheets are prepared by using 20 items from plankton tow sheet (circled numbers on the tow sheet), the plankton volumes and the data of the computer's calibration curve described above.

For the new Set I, a printout will include stations indicating errors or omissions of meter...
Figure 37.—Sample copies of standardization of data in sets for the first 10 stations of cruise 6607-J (see text for methods).
Figure 38.—Regression line derived from data of stations 60.50 to 80.55, Nos. 1 to 43 by order of occupancy on cruise 6607-J. The line was needed to obtain an estimated meter reading for station 60.70 on which a sample was collected and a proper set of wire angles recorded, but the tow was not repeated. When the line was drawn, the “reading”, 4.17 rev/sec, was obtained by taking the point off the line at the average tangent 1.102. All data derived from such an estimate are parenthesized in the data sets in Figure 37.
readings if any. A scatter diagram with regression data are printed out and a second and final Set I is printed with corrections showing estimated meter readings, volumes of water strained, and the SHF derived from them.

Note: A scatter diagram and regression data are prepared whether needed or not in order to present a visual record of meter behavior during a cruise.

The final step in standardizing the data is to multiply all counts of eggs and larvae from each plankton tow by the standardized haul factor (Fig. 30). The standardized counts are filed on species cards by cruise and station. Measured larvae are standardized by numbers per size to the nearest one-half millimeter, then grouped in size classes, e.g., Kramer (1971). Sardine eggs are standardized by stage of development and totaled in “Age in Days” (see Step IV in identification of eggs and larvae), e.g., Kramer (1971).

For ADP a coding system is used now to standardize the data. A number (code) is assigned to each fish larva, sized or unsized, as far as possible to order, family, genus or species (Fig. 31). The code numbers are added to the identifiers sheet as shown in Figure 32, and the key punch operator puts the following information on a card for each species: cruise number, station number, larva or egg code number, and the standard haul factor.

These data are then stored on tape for retrieval for analysis and/or publication.

**Data Publication**

The data for each year of surveys are compiled for publication in two series. The first is “Zooplankton volumes of the Pacific coast, . . . ,” the old Sets III (now Set II by ADP) for each survey, e.g., Thrailkill (1969). The second is “Sardine eggs and larvae and other fish larvae of the Pacific coast, . . . ,” which include the standardized haul factors for all stations occupied on each survey and positive occurrences of particular species as follows: Pacific sardine eggs by age in days, fish larvae by size classes including Pacific sardine, northern anchovy, jack mackerel and Pacific mackerel, and fish larvae unsized, including Pacific hake and rockfish spp., e.g., Kramer (1971).

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Sincere appreciation is extended to all personnel and organizations without whose cooperation the exacting procedures and extensive work of the CalCOFI could never be accomplished.

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