757



NOAA Technical Report NMFS SSRF- 757 A Profile of the Fish and Decapod Crustacean Community in a South Carolina Estuarine System Prior to Flow Alteration

Elizabeth Lewis Wenner, Malcolm H. Shealy, Jr., and Paul A. Sandifer

March 1982

U.S. DEPARTMENT OF COMMERCE National Oceanic and Atmospheric Administration National Marine Fisheries Service

5758 40

NOAA TECHNICAL REPORTS .

National Marine Fisheries Service, Special Scientific Report-Fisheries

The major responsibilities of the National Marine Fisheries Service (NMFS) are to monitor and assess the abundance and geographic distribution of fishery resources, to understand and predict fluctuations in the quantity and distribution of these resources, and to establish levels for optimum use of the resources. NMFS is also charged with the development and implementation of policies for managing national fishing grounds, development and enforcement of domestic fisheries regulations, surveillance of foreign fishing off United States coastal waters, and the development and enforcement of international fishery agreements and policies. NMFS also assists the fishing industry through marketing service and economic analysis programs, and mortgage insurance and vessel construction subsidies. It collects, analyzes, and publishes statistics on various phases of the industry.

The Special Scientific Report—Fisheries series was established in 1949. The series carries reports on scientific investigations that document long-term continuing programs of NMFS, or intensive scientific reports on studies of restricted scope. The reports may deal with applied fishery problems. The series is also used as a medium for the publication of bibliographies of a specialized scientific nature.

NOAA Technical Reports NMFS SSRF are available free in limited numbers to governmental agencies, both Federal and State. They are also available in exchange for other scientific and technical publications in the marine sciences. Individual copies may be obtained from D822, User Services Branch, Environmental Science Information Center, NOAA, Rockville, MD 20852. Recent SSRF's are:

722. Gulf menhaden, *Brevoortia patronus*, purse seine fishery: Catch, fishing activity, and age and size composition, 1964-73. By William R. Nicholson. March 1978, iii + 8 p., 1 fig., 12 tables.

723. Ichthyoplankton composition and plankton volumes from inland coastal waters of southeastern Alaska, April-November 1972. By Chester R. Mattson and Bruce L. Wing. April 1978, iii + 11 p., 1 fig., 4 tables.

724. Estimated average daily instantaneous numbers of recreational and commercial fishermen and boaters in the St. Andrew Bay system, Florida, and adjacent coastal waters, 1973. By Doyle F. Sutherland. May 1978, iv + 23 p., 31 figs., 11 tables.

725. Seasonal bottom-water temperature trends in the Gulf of Maine and on Georges Bank, 1963-75. By Clarence W. Davis. May 1978, iv + 17 p., 22 figs., 5 tables.

726. The Gulf of Maine temperature structure between Bar Harbor, Maine, and Yarmouth, Nova Scotia, June 1975-November 1976. By Robert J. Paw-lowski. December 1978, iii + 10 p., 14 figs., 1 table.

727. Expendable bathythermograph observations from the NMFS/MARAD Ship of Opportunity Program for 1975. By Steven K. Cook, Barclay P. Collins, and Christine S. Carty. January 1979, iv + 93 p., 2 figs., 13 tables, 54 app. figs.

728. Vertical sections of semimonthly mean temperature on the San Francisco-Honolulu route: From expendable bathythermograph observations, June 1966-December 1974. By J. F. T. Saur, L. E. Eber, D. R. McLain, and C. E. Dorman. January 1979, iii + 35 p., 4 figs., 1 table.

729. References for the identification of marine invertebrates on the southern Atlantic coast of the United States. By Richard E. Dowds. April 1979, iv + 37 p.

730. Surface circulation in the northwestern Gulf of Mexico as deduced from drift bottles. By Robert F. Temple and John A. Martin. May 1979, iii + 13 p., 8 figs., 4 tables.

731. Annotated bibliography and subject index on the shortnose sturgeon, Acipenser brevirostrum. By James G. Hoff. April 1979, iii + 16 p.

732. Assessment of the Northwest Atlantic mackerel, Scomber scombrus, stock. By Emory D. Anderson. April 1979, iv + 13 p., 9 figs., 15 tables.

733. Possible management procedures for increasing production of sockeye salmon smolts in the Naknek River system, Bristol Bay, Alaska. By Robert J. Ellis and William J. McNeil. April 1979, iii + 9 p., 4 figs., 11 tables.

734. Escape of king crab, *Paralithodes camtschatica*, from derelict pots. By William L. High and Donald D. Worlund. May 1979, iii + 11 p., 5 figs., 6 tables.

735. History of the fishery and summary statistics of the sockeye salmon, Oncorhynchus nerka, runs to the Chignik Lakes, Alaska, 1888-1956. By Michael L. Dahlberg. August 1979, iv + 16 p., 15 figs., 11 tables.

736. A historical and descriptive account of Pacific coast anadromous salmomid rearing facilities and a summary of their releases by region, 1960-76. By Roy J. Wahle and Robert Z. Smith. September 1979, iv + 40 p., 15 figs., 25 tables.

737. Movements of pelagic dolphins (*Stenella* spp.) in the eastern tropical Pacific as indicated by results of tagging, with summary of tagging operations, 1969-76. By W. F. Perrin, W. E. Evans, and D. B. Holts. September 1979, iii + 14 p., 9 figs., 8 tables.

Environmental baselines in Long Island Sound, 1972-73. By R. N. Reid,
A. B. Frame, and A. F. Draxler. December 1979, iv + 31 p., 40 figs., 6 tables.

739. Bottom-water temperature trends in the Middle Atlantic Bight during spring and autumn, 1964-76. By Clarence W. Davis. December 1972, iii + 13 p., 10 figs., 9 tables.

740. Food of fifteen northwest Atlantic gadiform fishes. By Richard W. Langton and Ray E. Bowman. February 1980, iv + 23 p., 3 figs., 11 tables.

741. Distribution of gammaridean Amphipoda (Crustacea) in the Middle Atlantic Bight region. By John J. Dickinson, Roland L. Wigley, Richard D. Brodeur, and Susan Brown-Leger. October 1980, vi + 46 p., 26 figs., 52 tables.

742. Water structure at Ocean Weather Station V, northwestern Pacific Ocean, 1966-71. By D. M. Husby and G. R. Seckel. October 1980, 18 figs., 4 tables.

743. Average density index for walleye pollock, *Theragra chalcogramma*, in the Bering Sea. By Loh-Lee Low and Ikuo Ikeda. November 1980, iii + 11 p., 3 figs., 9 tables.



NOAA Technical Report NMFS SSRF- 757

A Profile of the Fish and Decapod Crustacean Community in a South Carolina Estuarine System Prior to Flow Alteration

Elizabeth Lewis Wenner, Malcolm H. Shealy, Jr., and Paul A. Sandifer

March 1982

U.S. DEPARTMENT OF COMMERCE Malcolm Baldrige, Secretary National Oceanic and Atmospheric Administration John V. Byrne, Administrator National Marine Fisheries Service William G. Gordon, Assistant Administrator for Fisheries The National Marine Fisheries Service (NMFS) does not approve, recommend or endorse any proprietary product or proprietary material mentioned in this publication. No reference shall be made to NMFS, or to this publication furnished by NMFS, in any advertising or sales promotion which would indicate or imply that NMFS approves, recommends or endorses any proprietary product or proprietary material mentioned herein, or which has as its purpose an intent to cause directly or indirectly the advertised product to be used or purchased because of this NMFS publication.

CONTENTS

Introduction
Study area
Materials and methods
Data collection
Data analysis
Results
Hydrographic parameters
Community composition and diversity
Temporal and spatial distributions — fishes
Temporal and spatial distributions — decapods
Biomass estimates
Discussion
Acknowledgments
Literature cited

Figures

1.	Station locations in North and South Santee Rivers, S.C.	2
2.	Monthly fluctuations in number of species and number of individuals (log transformed) of fishes and decapod crustaceans at sampling sites in the North and South Santee Rivers, 1975-76	6
3.	Two-way coincidence tables of constancy and fidelity which compare species associations with stations in the North and South Santee Rivers, 1975-76. The species associations, designated alphabetically, resulted from cluster analysis of species (dendrogram not shown) collected from the Santee system. Species comprising these	
	associations are listed in Table 5	7
4.	Abundance of A) <i>Micropogonias undulatus</i> and B) <i>Anchoa mitchilli</i> by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total	
	number of collections at a station	9
5.	Abundance of A) <i>Ictalurus catus</i> and B) <i>Bairdiella chrysoura</i> by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total number of collections at a station	10
6.	Abundance of <i>Trinectes maculatus</i> by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total number of collections at a station	11
7.	Seasonal abundance and length-frequency distribution of <i>Penaeus setiferus</i> within the North and South Santee Rivers. Ratios over the bars indicate number of collections where shrimp were captured to total number of col-	
	lections at a station	12
8.	Abundance of A) <i>Penaeus aztecus</i> and B) <i>Callinectes sapidus</i> . Ratios over the bars indicate number of collections where shrimp were captured to total number of collections at a station	13

Tables

1.	Total number and total weight of fishes collected from 1975 and 1976 in estuaries of the North and South Santee Rivers, S.C. Species are listed in order of abundance, and data are pooled from the 2-yr sampling	
	period.	4
2.	Total number and total weight of decapod Crustacea collected from 1975 and 1976 in the North and South	
	Santee Rivers. Species are listed in order of abundance and data are pooled over the 2-yr sampling period	5
3.	Correlation between $\ln (n + 1)$ transformed values of number of species and number of individuals of fishes	
	and decapods in relation to environmental factors	6
4.	Groups formed from cluster analysis of species of fishes and decapod Crustacea collected in the North and	
	South Santee Rivers from 1975 and 1976	7
5.	Estimates of density and number of individuals/ha for fishes caught by trawls from estuaries along the Gulf and	
	east coasts of the United States	12

A Profile of the Fish and Decapod Crustacean Community in a South Carolina Estuarine System Prior to Flow Alteration¹

ELIZABETH LEWIS WENNER, MALCOLM H. SHEALY, JR., and PAUL A. SANDIFER²

ABSTRACT

The seasonal distribution and abundance of fishes and decapod Crustacea collected by 6 m otter trawl from the North and South Santee Rivers, South Carolina, were examined over a 2-year sampling period. Species richness was greatest during summer and at stations located in proximity to the river mouths. Although species richness was found to be related to salinity, temperature, depth, and dissolved oxygen, it was most noticeably affected by a spring freshet which considerably lowered richness and abundance.

Eleven species accounted for 93% of the number and $\sim 70\%$ of the total fish biomass taken in both rivers: Micropogonias undulatus, Anchoa mitchilli, Bairdiella chrysoura, Stellifer lanceolatus, Ictalurus catus, Cynoscion regalis, Dorosoma petenense, Leiostomus xanthurus, Trinectes maculatus, Brevoortia tyrannus, and Symphurus plagiusa. White shrimp, Penaeus setiferus; brown shrimp, P. aztecus; and blue crabs, Callinectes sapidus, comprised over 96% by number and weight of the decapod fauna collected in both rivers. Dominant fishes were present in fairly equal abundance throughout the year and utilized the Santee system as either a residential or nursery area, while P. setiferus and P. aztecus were more seasonal in their pattern of appearance and abundance.

Length-frequency analysis showed the Santee system fish fauna to be composed mostly of juvenile specimens. Their presence throughout the year indicated that the Santee is a temporally stable and relatively nonstressed system and an important nursery area.

The predominance of juveniles accounted for lower biomass (kg/ha) of fishes in the Santee system compared to values for other estuaries along the Atlantic coast of the United States. The continued importance of juvenile fishes and shrimp in the Santee system is questionable in view of salinity changes in the nursery habitat following proposed river rediversion.

INTRODUCTION

A number of published accounts are available on community ecology of estuarine fauna along the northeastern and southeastern coasts of the United States. Despite this, few comprehensive studies exist on distributional patterns and faunal composition of estuarine megafauna, such as fishes and decapods, from a system which has experienced as many manipulations as the Santee River, S.C. The drainage basin of the Santee encompasses 41,000 km² in North and South Carolina. The Santee was the fourth largest river on the U.S. east coast prior to diversion of most of its flow into the Cooper River in 1942. Diversion not only lowered the annual mean discharge from 525 m³/s to 74 m³/s but also caused severe shoaling in Charleston Harbor at the mouth of the Cooper River (Kjerfve and Greer 1978). Changes in the amount of freshwater flow completely altered the supply and deposition of sediments, erosion patterns, salinity regime, flooding characteristics, and floral and faunal communities (Kjerfve 1976). After diversion, the salinity in the Santee distributaries, the North and South Santee Rivers, increased sharply. In addition, large quantities of fine-grained suspended sediments were transported into the Cooper River and, eventually, into Charleston Harbor.

The costly necessity to dredge Charleston Harbor continuously prompted a rediversion project begun in 1975 whereby 80% of the Cooper River flow eventually will be directed back into the Santee system (Kjerfve 1976). Upon projected completion of rediversion on the North and South Santee Rivers is purely 428 m3/s (Kjerfve and Greer 1978). Although the impact of rediversion on the North and South Santee Rivers is purely speculative, it is likely to reduce the net salinity in the Santee system and increase amounts of fine-grained suspended sediments. Kjerfve and Greer (1978) cautioned that these combined changes may ultimately end the economically important American oyster, Crassostrea virginica, and hard clam, Mercenaria mercenaria, fisheries flourishing at present in this river. The effect of rediversion on fishes and decapod Crustacea also may be substantial. Shealy and Bishop (1979) suggested that population changes in penaeid shrimp may occur, and the extent of nursery areas may be affected. Fishes such as Ictalurus catus, I. punctatus, Morone saxatilis, and Anguilla rostrata, which are frequently encountered in lower salinity regions of estuaries, may increase in abundance in the Santee system; however, the effect on most species, such as the euryhaline sciaenid fishes which are numerically dominant in South Carolina estuaries (Shealy et al. 1974), cannot be predicted.

The present study examines quantitative annual and seasonal variability, diversity, and species assemblages of fishes and decapod Crustacea found in the channel of the lower North and South Santee Rivers. Our primary consideration is to describe the megafaunal community as it currently exists and to relate distributional patterns to abiotic factors which may influence the community after rediversion.

STUDY AREA

All sampling stations were located within the Santee River system (Fig. 1). The Santee River provides the major headwaters

¹Contribution No. 139 from the South Carolina Marine Resources Center. ²Marine Resources Research Institute, P.O. Box 12559, Charleston, SC 29412.



Figure 1.-Station locations in North and South Santee Rivers, S.C.

supplying Lakes Marion and Moultrie. About 23 km from the ocean, the Santee River bifurcates to form the North and South Santee Rivers.

The Santee system has been classified by Kjerfve and Greer (1978) as a partially mixed estuary with weak-to-moderate salinity stratification and gravitational circulation. However, this classification is variable due to tidal fluctuations as well as variations in saltwater intrusion and freshwater discharge (Cummings 1970³; Stephens et al. 1975; Kjerfve 1976; Nelson 1976; Burrell 1977; Calder et al. 1977). The South Santee River receives less freshwater drainage, with the result that saltwater intrusion is greater than in the North Santee River.

The two distributaries differ somewhat with regard to bathymetry since the North Santee River is slightly deeper than the South Santee River. Substrate in both rivers is very similar, being predominately coarse to fine-grained sand and shell of oceanic origin at the mouths, and hard mud and sand mix in the intermediate reaches of the estuary, replaced by fine-grained sand of inland origin in the upper estuary (Calder et al. 1977).

Dissolved oxygen values fluctuate seasonally, being usually 9-14 mg/liter in winter and \geq 4 mg/liter in summer (Cummings footnote 3; Nelson 1976; Mathews⁴).

MATERIALS AND METHODS

Data Collection

We sampled eight stations which were located in the channel at 1, 4, 7, and 11 river miles from the mouths of the North and South Santee Rivers (Fig. 1). Hereafter, we will refer to these stations as NS01, NS04, NS07, and NS11 in the North Santee River and SS01, SS04, SS07, and SS11 in the South Santee River. Stations were sampled monthly over a 2-yr period from January 1975 through December 1976, with the following exceptions which were not included in our analysis: SS11 was not sampled in 1975; NS11 was sampled with a 5 m (16 ft) trawl in May and July 1975; and NS07 was not successfully trawled in May 1975.

All collections were made with a 6 m (12 ft) semiballoon otter trawl with 8 m headrope, composed of 2.5 cm (1 in) stretch mesh throughout. A complete description of the trawl is given by Shealy et al. (1974). Twenty-minute tows were made against floodtide during daylight hours at a speed of 1.3 m/s (2.5 kn), which resulted in a coverage of 1.5 ± 0.4 km during a tow.

Bottom-water samples were collected with 6 liter capacity Van Dorn bottles 0.3 m above the bottom at each station prior to trawling. Water temperature was read from stem thermometers mounted within the Van Dorn bottles. Salinity was measured in the laboratory with a Beckman RS7B induction salinometer. Dissolved oxygen was determined by the Winkler-Carpenter method (Strickland and Parsons 1968). Turbidity was determined with a Hach Model 2100A turbidimeter. Specimens were either processed in the field or preserved in 10% Formalin and returned to the laboratory for identification, measuring, and weighing. All specimens were weighed to the nearest 0.1 g and counted. We also recorded measurements (total length for fishes, carapace width for crabs, and total length for shrimps) for all species numbering ≤ 50 specimens per tow. At stations where the trawl caught larger numbers of organisms, we subsampled the catch as follows: If ≥ 50 to ≤ 250 individuals were collected, then a minimum of 50 randomly selected specimens were measured; if > 250 to ≤ 500 individuals were caught, then

³Cummings, T. R. 1970. A reconnaissance of the Santee River estuary, South Carolina. A report prepared by United States Geological Survey, Water Resources Division, Columbia, S.C., 96 p.

⁴T. Mathews, Assistant Marine Scientist, South Carolina Marine Resources Research Institute, Charleston, SC 29412, pers. commun. December 1979.

20% of the catch was measured; when >500 were caught, 10% of the catch was measured.

Data Analysis

I

Cluster analysis was used to define assemblages of fishes and decapod crustaceans and to determine degree of similarity among stations. Prior to cluster analysis, data were logarithmically transformed by $\log_{10} (x + 1)$, where x is number of individuals for a given species. We reduced data by elimination of species which occurred in fewer than three collections during a sampling period and by elimination of collections which contained only one species.

The methods of cluster analysis used are described in detail by Boesch (1977). The Bray-Curtis coefficient (Clifford and Stephenson 1975) was used to compute similarity values. Symmetrical similarity matrices were computed for both the North and South Santee Rivers on data from the 2-yr sampling period with collections as entities and species as attributes (normal analysis), and with species as entities and sites as attributes (inverse analysis). Entities were classified into related groups by using flexible sorting (Lance and Williams 1967) with $\beta =$ -0.25.

Two separate dendrograms were generated for each river: A dendrogram which indicated association of all collections during the 2-yr sampling period based on their faunal content and a dendrogram which indicated association of all species from the collections made during the 2-yr sampling period. We used postclustering techniques of nodal analysis (Williams and Lambert 1961; Lambert and Williams 1962) to examine species and station coincidences. Nodal analysis diagrams were made by using patterns of constancy (a measure of how consistently a species is found in a site group) and fidelity (a measure of how restricted a species is to a site group).

An index of abundance (Musick and McEachran 1972; Elliott 1977) was used to compare numbers and weights of selected dominant species and is expressed as:

ndex of Abundance =
$$\frac{1}{n} \sum_{l=1}^{n} \log_{10} (x + 1),$$

where x = number or weight of individuals of a given species and n = number of collections in a chosen time frame.

We determined biomass and density estimates for fishes and decapods from computations of area swept for trawl gears. Estimates of area swept (*a*) were determined by the following equation given by Roe (1969):

$$a = \frac{K \times M \times (0.6 \text{ H})}{10,000 \text{ m}^2/\text{ha}}$$

where K is speed in meters per hour, M is time in hours fished, and H is headrope length in meters (Klima⁵). Roe (1969) assumed an effective swath of about 60% of the headrope length as established by Wathne (1959). The area swept by our 6 m otter trawl was estimated to be 0.72 ha/tow based on the method described by Roe (1969).

Hydrographic Parameters

Bottomwater temperatures were very similar between rivers and among stations. Temperatures were lowest in both the North and South Santee Rivers during February and March 1975 and January and February 1976. In the North Santee River, temperatures gradually increased from April to reach a peak in either August (1976) or September (1975). The warmest month in the South Santee River during both years of sampling was August. Based on temperature over the 2-yr sampling period, winter encompassed January, February, and March; spring, April, May, and June; summer, July, August, and September; and fall, October, November, and December.

Salinities were extremely variable both seasonally and among stations. Freshwater outflow increased in the Santee watershed from 14.2 m³/s to an average of 679.3 m³/s between mid-March and mid-April 1975 and to 238.5 m3/s from mid-May until late June 1975 (Burrell 1977). These freshets considerably lowered salinities at stations in both rivers. Salinities were also variable in 1976 but the extreme fluctuations caused by freshwater outflow were not as evident as in 1975. Except during periods of high runoff when freshwater was found throughout the system, salinity decreased from stations located at the river mouth to those located upstream. Salinities at stations SS01 and NS01 ranged from 0.2 to 32.9%, which characterized these stations as limnetic-euhaline by the Venice System (Symposium on the Classification of Brackish Waters 1958). Stations SS04 and NS04 were limnetic-polyhaline (0.1-26%), while SS07 and NS07 were limnetic-mesohaline (0.1-15.9%). Salinities at NS11 and SS11 ranged from <0.1 to 1.4% and were within the limnetic-oligohaline salinity regime.

Community Composition and Diversity

Eighty species of fishes were collected from the South Santee River and 64 species from the North Santee River during the 1975-76 sampling period (Table 1). Eleven species accounted for 93% of the total number of specimens and 70% of the total fish biomass taken in both rivers: Atlantic croaker, *Micropogonias* undulatus; bay anchovy, *Anchoa mitchilli*; silver perch, *Bairdiella chrysoura*; star drum, *Stellifer lanceolatus*; white catfish, *Ictalurus catus*; weakfish, *Cynoscion regalis*; threadfin shad, *Dorosoma petenense*; spot, *Leiostomus xanthurus*; hogehocker, *Trinectes maculatus*; Atlantic menhaden, *Brevoortia tyrannus*; and blackcheek tonguefish, *Symphurus plagiusa*. In both rivers, *M. undulatus* was the most abundant species collected. With regard to biomass, however, *M. undulatus* was outranked by *I. catus* in the North Santee River and Bairdiella chrysoura in the South Santee River.

The decapod crustaceans were represented by 22 species in the North Santee River and 18 species in the South Santee River. Although fewer species of decapods than fish were collected, the decapods dominated in terms of total number of individuals captured (Table 2). The numerical dominance of the decapods was due to extremely large catches of the white shrimp, *Penaeus setiferus*, especially in the South Santee River. This species was by far the most abundant decapod collected in both rivers and also dominated other decapods in terms of biomass. *Penaeus setiferus*, together with the brown shrimp, *P. aztecus*, and the blue crab, *Callinectes sapidus*, comprised over 96% by number and weight of the total decapod fauna collected in both rivers.

³Klima, E. F. 1976. A review of the fishery resources in the western central Atlantic. West. Cent. Atl. Fish. Comm. Publ. 3, 77 p.

Table 1.-Total number and total weight (kg) of fishes collected from 1975 and 1976 in estuaries of the North and South Santee Rivers, S.C. Species are listed in order of abundance, and data are pooled for the 2-yr sampling period.

Ophidion marginatum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.172 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.016 Selene vomer 3 0.02 0.013 0.01 Lutarus griesus 4 0.03 0.041 Membras marinica 2 0.01 0.005 <0.01 Morone saxatilis 3 0.02 0.054 Eucinostomus argenteus 2 0.01 0.005 <0.01 Arius felis 3 0.02 0.041 Gobionellus hastatus 2 0.01 0.093 0.05 Eucinostomus sp. 3 0.02 0.042 Hypsobleninus ionthas 2 0.01 0.007 <0.01 Prionotus scitulus 3 0.02 0.007 Chaetodipterus faber 1 0.01 0.022 0.01 Elettris pisonis 2 0.01 0.014 Archosargus probatocephalus 1 0.01 0.033 <th>Species</th> <th>Total no.</th> <th>Percent of total catch</th> <th>Total weight</th> <th>Percent weight</th> <th>Species</th> <th>Total no.</th> <th>Percent of total catch</th> <th>Total weight</th> <th>Percen weight</th>	Species	Total no.	Percent of total catch	Total weight	Percent weight	Species	Total no.	Percent of total catch	Total weight	Percen weight
Micropone unidation 3.35 23.68 20.22 0.102 Micropone unidation 4.365 11.7 23.84 Returns ratis 1.97 13.33 5.8.429 29.16 Baidelise drysoura 2.187 15.8.3 3.54 Returns ratis 1.97 13.33 5.8.429 29.16 Baidelise drysoura 6.33 4.34 3.44 Brevoring (rpumine 5.3 5.32 1.800 6.9 K. Lubara ratis 6.3 4.33 2.34 Brevoring (rpumine 5.31 3.53 2.60 1.50 Cronician ratis 3.64 2.20 1.50 Decomp prime 4.18 2.30 5.517 2.69 Brevoring (rpumine 3.64 2.19 1.50 Statististististististististististististist	forth Santee River:					South Santee River:				
Timesis machina 1,23 21.66 15.21 7.61 Anchos michili 2,447 15.22 33.547 Archos michili 1,932 10.06 2.385 1.44 Stellfer lancedatas 6.15 4.40 33.147 Devicorite framme 843 5.32 1.50 1.51 2.51 2.51 2.51 2.54 2.55 2.56 2.	Micropogonias undulatus	3,535	23.68	20.282	10.12		4,385	31.72	23.815	13.9
Accha michili 1,902 10.06 2.395 1.44 Stafifer lancealus 645 4.40 3.440 Stafifer lancealus 766 5.13 3.61 1.82 Conscion regula 595 4.32 2.743 Derosoma preference 401 2.89 3.03 2.745 2.702 Baridelli debyson 3.11 3.56 8.621 4.30 Leintomus aunthana 3.84 2.20 1.103 Derosoma preference 4.01 3.15 4.56 Dynaphina preference 2.02 1.123 Derosoma preference 4.01 1.00 1.03 1.51 4.56 Dynaphina preference 2.01 1.123 Calabra punctiona 1.12 0.75 3.88 Opamas tan 85 0.61 7.75 Arak / 60 4.03 3.03 0.17 Paralchysis lethosis netziz 38 0.23 1.342 Arak / 60 4.03 3.03 0.17 Paralchysis lethosis netziz 38 0.03 0.03 0.03 0.03	Trinectes maculatus	3,233	21.66	15.251	7.61		2,944	21.29	4.259	2.4
Brevoor reguls 64 5.2 13.60 6.79 <i>Laulang catus</i> 0.25 4.30 2.2478 Conscion reguls 555 3.72 2.58 1.26 Conscions peteres 400 2.378 Darcosona peterence 407 3.13 3.68 3.63 4.30 2.102 1.139 Darcosona peterence 407 3.13 3.81 0.91 Previous in seminar 304 2.20 1.139 Lessionas samhara 20 1.39 9.135 4.56 Symphurs placing 20 1.139 2.139 Lessionas samhara 20 1.39 9.135 4.56 Appandensia contha 60 40 0.137 Frankchrys denotines 1.13 1.38 1.564 7.52 Hyposhemis contha 60 40 0.254 0.734 1.564 7.52 Hyposhemis contha 60 0.222 1.535 Praikchrys denotine 23 0.17 0.38 0.042 Contropersits frain 20 0.145 0.224	Ictalurus catus	1,975	13.23	58.429	29.16	Bairdiella chrysoura	2,187	15.82	33.547	19.5
Solijor immenolnia 766 5.13 3.61 1.82 Conscion regula 995 4.30 2.478 Bandelle drygourn 531 3.55 3.62 4.30 Laiotomus zunthman 344 2.20 9.100 Decomom petromes 477 3.13 8.43 2.40 1.457 Somplumes plasma 211 1.30 9.13 4.30 3.13 1.31 1	Anchoa mitchilli	1,502	10.06	2.895	1.44	Stellifer lanceolatus	636	4.60	3.419	2.0
Conscion regile 55 3.72 2.52 1.26 Dorasome prevenes 400 2.102 Bridelic dryspun 51 3.56 K.51 4.50 Leistome samthans 344 2.20 11.03 Dorasome prevenes 467 3.13 1.81 0.91 Trinecter maculans 344 2.20 1.57 Symphurs plagins 201 1.39 9.33 4.56 Symphurs plagins 231 1.12 1.21 Leasterne purchass 1.10 0.82 1.78 Paralechysic it yamms 240 0.01 7.79 Marchark purchass 1.20 0.33 0.37 Opanas tau 80 0.61 7.79 Marchark purchass 1.03 0.51 0.52 Opanas tau 80 0.61 7.79 Marchark purchass 25 0.17 0.53 Opanas tau 80 0.62 0.22 1.032 Paralchys detations 25 0.17 0.54 Conscionaria tauta 29 0.21 0.62	Brevoortia tyrannus	824	5.52	13.600	6.79	Ictalurus catus	625	4.52	25.443	14.8
Bandal chynaum 31 3.56 8.62 4.00 Leistama maruharus M4 2.32 9.00 Dornsome petrones 467 3.13 1.81 0.91 Triceste maruharus 104 2.30 1.135 Symphunse playaa 418 2.40 5.37 2.99 Erotoris tyranus 230 1.9 2.10 1.135 Erallarus furcitais 142 0.53 1.81 Paralichthys ichosingma 131 0.84 0.73 1.38 Opmans tau 83 0.01 7.73 3.84 Opmans tau 83 0.02 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.12 0.02 0.12 0.02 0.12 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03 0.03	Stellifer lanceolatus	766	5.13	3.651	1.82	Cynoscion regalis	595	4.30	2.478	1.4
Beindheil obygoune 51 3.56 8.62 4.30 Leistomus sunturus Ma 2.32 9.100 Decomma peterines 461 3.13 8.81 2.69 Inconstria frammus 291 2.10 11.039 Leistomus sunturus 130 3.23 5.85 Symphunes plasias 230 1.9 2.13 1.32 1.32 1.32 1.32 1.33 1.33 1.35 5.55 Symphunes plasias 1.30 1.32 1.32 1.32 1.32 1.32 1.32 1.32 1.32 1.33 1.34 1.34 1.34 0.15 1.32 0.13 0.34 0.15 1.33 0.02 1.33 0.14 1.34 0.35 0.23 0.33 0.17 0.40 0.42 0.03 0.23 1.33 0.15 Characon neuclass 3.0 0.23 0.33 0.23 0.33 0.23 0.33 0.23 0.33 0.23 0.33 0.23 0.33 0.23 0.33 0.23 0.33	Cynoscion regalis	555	3.72	2,528	1.26	Dorosoma petenense	400	2.89	2.102	1.2
Symphanes phatisan 418 2.80 5.37 2.69 Personsite pramas 291 2.10 1.109 Leatomus punctata 150 1.00 3.623 1.81 Paralichtys ichostigem 155 1.12 13.74 Leatomus punctata 150 1.00 3.623 1.81 Paralichtys ichostigem 15 0.43 0.31 Peralichtys ichostigem 131 0.88 15.664 7.82 Hypobhemias ionta 60 0.43 0.187 Minisrithus americans 50 0.33 0.33 0.17 Paralichtys ichitus americans 34 0.22 1.033 Paralichtys ichitus 25 0.17 0.03 Cynoxion nebuloas 30 0.22 1.033 Paralichtys ichitus 25 0.17 0.04 Corporatis strata 28 0.19 0.030 0.51 Appular ichitus ichitus 25 0.17 0.13 0.16 Advas sactualis 0.16 0.15 Ciprostis strata 29 0.14 0.05 0.15	Bairdiella chrysoura	531	3.56	8.621	4.30	Leiostomus xanthurus	348	2.52	9.100	5.3
Lansan sumhana 207 1.39 9.13 4.56 Symphane pagasa 220 1.99 2.193 Leadnars purchas 150 1.00 3.623 1.81 Paralichtys lehnings in this 155 1.12 1.324 Leadnars purchas 10 0.85 7.785 3.8 Opsames tau 80 0.43 0.134 Menitchthas americanus 57 0.8 0.53 0.37 Paralichtys lehnings 0.42 0.246 Opsames tau 20 0.13 0.33 0.37 Chroscom nebulosas 30 0.22 1.033 Paralichtys lehning 25 0.17 0.348 0.04 Corroysitis striata 28 0.30 0.031 Paralichtys lehning 1.10 0.15 0.16 0.032 0.01 0.032 0.01 0.033 0.01 0.033 0.01 0.032 0.01 0.032 0.01 0.032 0.01 0.032 0.01 0.032 0.01 0.033 0.01 0.033 0.01	Dorosoma petenense	467	3.13	1.831	0.91	Trinectes maculatus	304	2.20	1.657	0.9
Induits 100 3.62 1.81 Paralechtys fednarigena 155 1.12 1.27 Paralchhys lefhatigena 131 0.88 15.66 7.32 3.84 Oppanis fun 85 0.61 7.430 Paralchhys lefhatigena 131 0.88 15.66 7.32 84 0.33 0.33 0.34 0.351 Meticirhus americanus 34 0.25 0.346 Urophysis regia 29 0.19 1.320 0.75 Cynascion rebulatus 39 0.22 1.032 Paralchhys dentaire 23 0.17 0.340 Cynascion rebulatus 23 0.18 0.032 Angualla rotaria 19 0.13 0.271 1.130 0.033 3.30 3.30 0.12 1.131 0.041 0.072 0.041 0.071 0.033 3.30 1.12 0.130 0.333 3.30 1.12 0.130 0.130 0.131 0.130 0.131 0.131 0.130 0.131 0.130 0.130 0.130 <td>Symphurus plagiusa</td> <td>418</td> <td>2.80</td> <td>5.387</td> <td>2.69</td> <td>Brevoortia tyrannus</td> <td>291</td> <td>2.10</td> <td>11.039</td> <td>6.4</td>	Symphurus plagiusa	418	2.80	5.387	2.69	Brevoortia tyrannus	291	2.10	11.039	6.4
International 100 3.62 1.81 Parallechtys lehonizyma 1.12 1.74 Frankchhys lehonizyma 131 0.88 15.66 7.82 Mysoobennis kontas 60 0.43 0.137 Frankchhys lehonizyma 131 0.88 0.531 <td< td=""><td>Leiostomus xanthurus</td><td>207</td><td>1.39</td><td>9.135</td><td>4.56</td><td>Symphurus plagiusa</td><td>220</td><td></td><td></td><td>1.2</td></td<>	Leiostomus xanthurus	207	1.39	9.135	4.56	Symphurus plagiusa	220			1.2
Leaharas 142 0.95 7.85 3.88 Opname iau 85 0.61 7.40 Peralichtys lehnistioning 13 0.38 15.64 7.82 Hypsoblemiss hentiz 58 0.43 0.187 Menticrhus americanus 57 0.33 0.33 0.31 0.56 Hypsoblemiss hentiz 42 0.03 1.502 Arius fels 49 0.33 1.040 0.51 Menticrhus americanus 44 0.25 0.23 Peralichtys lehnizus 25 0.17 0.050 0.04 Urphysis regin 28 0.20 0.33 Ansu spidistram 17 0.11 0.52 0.29 Peralichtys lehnizus 25 0.18 0.076 Cripnics cargin 16 0.11 2.52 0.29 Perplenis triacanthus 25 0.18 0.076 Coprinus cargin 10 0.111 0.525 0.29 Perplenis disachthis 30 0.14 0.085 Coprinus cargin 0.035 0.13 <td< td=""><td>Ictalurus punctatus</td><td>150</td><td>1.00</td><td>3.623</td><td>1.81</td><td></td><td>155</td><td></td><td></td><td>7.7</td></td<>	Ictalurus punctatus	150	1.00	3.623	1.81		155			7.7
Parallechtops lehonizigna 131 0.88 15.66 7.82 Ipproblemins kentli 58 0.43 0.36 Mentiorthu americanus 57 0.38 0.33 0.36 Hypoblemins kentli 58 0.42 0.36 Arias Jolis 49 0.33 0.132 0.51 Menticrhus americanus 34 0.25 1.034 Oparatus fan 29 0.19 1.034 0.42 Conscion nebuloss 30 0.22 1.033 Apaulichnys fandin 25 0.17 0.43 0.23 0.621 0.627 Eropus crossona 25 0.17 0.13 0.232 Pophycis finita 23 0.17 0.130 Cynixis carpio 16 0.11 2.393 1.39 Chiorascombras chysuns 21 0.15 0.160 Cynixis carpio 16 0.11 2.393 1.39 Chiorascombras chysuns 21 0.14 0.36 Cynixis carpio 16 0.11 2.393 1.40 0.44 0.3	Ictalurus furcatus	142	0.95	7.785	3.88		85			4.3
Manicembra americanus 57 0.38 0.31 0.26 Prinsobiennis henria 58 0.42 0.43 Arius feis 49 0.33 1.034 0.51 Menicicrhus americanus 44 0.23 0.34 Arius feis 49 0.33 1.042 0.51 Menicicrhus americanus 44 0.23 0.33 Paralichhys dentatis 25 0.17 0.040 0.040 Compriss fraita 29 0.21 0.033 Aloss sapidistina 17 0.11 0.13 2.01 1.5 Perplika tiraunthus 25 0.17 0.13 Aloss sapidistina 17 0.11 0.282 0.29 Perplika singulotistina 19 0.14 0.076 Chronscombras chrystura 13 0.09 0.022 0.16 Alosa sapidistina 19 0.14 0.036 Chronscombras chrystura 18 0.01 0.022 0.01 Alcoas sapidistina 19 0.14 0.036 Chronscombras chrystura 18 0.03		131	0.88							0.1
Ucaphyse regia 90 0.33 0.39 0.17 Paralechtys derivans 14 0.35 0.324 Opamis fin 29 0.19 1.034 0.75 Crossion relations 34 0.22 1.034 Paralechtys dentaire 25 0.17 0.480 0.42 Crossion relations 29 0.31 Apaille rotation 25 0.17 0.480 2.09 Propress regia 28 0.20 0.33 Apaille rotation 10 0.13 0.230 Chlorascombras 23 0.17 0.130 Cyprise fordina 17 0.11 0.182 Opamis 0.132 0.13 0.130 Cyprise fordina 13 0.09 0.225 0.16 Alona serivisis 21 0.14 0.130 Consoin rebuloas 11 0.07 0.047 0.05 Erropia crossoin 19 0.14 0.046 Chorascherbuloas 11 0.07 0.047 0.031 Darais subia 10.12 0.043										0.1
Arias fols 49 0.33 1.024 0.51 Menticima annexama 34 0.25 1.033 Paralicho's dintatus 25 0.17 0.050 0.755 Conscion achulosa 30 0.22 1.033 Paralicho's dintatus 25 0.17 0.075 Conscion achulosa 26 0.19 0.331 Ansulia contrata 19 0.13 2.301 1.15 Paprilis timaanthas 26 0.19 0.331 Absus sapidisami 17 0.11 0.141 0.066 0.0664xos stimantas 21 0.17 0.196 Conocol metulosati 13 0.09 0.022 0.16 Aloss apticisamin 19 0.14 0.156 Conocol metulosati 12 0.08 0.032 0.16 Aloss apticisamin 19 0.14 0.136 Conocol metulosati 12 0.08 0.035 0.12 Calaras tynicisamin 10 0.12 0.421 0.421 0.421 0.421 0.421 0.421 0.421										0.8
Openant tatu 29 0.19 1.92 0.75 Cproscion nebulacia 30 0.22 1.03 Erropis crossitut 25 0.17 0.40 0.207 (netropricit siriant 28 0.20 0.51 Erropis crossitut 25 0.17 0.40 Urophycis fordiant 23 0.17 0.10 Alos augulassitut 10 0.11 0.23 0.22 0.13 0.17 0.10 Conscion nebulassit 13 0.17 0.13 0.16 0.16 0.17 0.19 Conscion nebulassit 13 0.09 0.023 0.01 Alosa astrivisit 20 0.14 0.019 Conscion nebulassit 13 0.09 0.023 0.01 Alosa astrivisit 20 0.14 0.026 Conscion nebulassi 13 0.09 0.023 0.015 Alosa astrivisit 20 0.14 0.026 Conscion nebulassi 13 0.17 0.18 0.015 0.015 0.015 0.015 0.015										0.8
Prinkehovs denatus 25 0.17 0.404 0.42 Centropristi striana 29 0.21 0.627 Anguila rostrata 19 0.13 2.301 1.15 Peprilss triananthas 26 0.19 0.351 Anguila rostrata 19 0.11 0.14 0.66 0.66bexx strumous 21 0.17 0.130 Constructionantial 10 0.11 0.382 0.29 Peprilss eligiptions 21 0.15 0.196 Construction relulosats 13 0.09 0.22 0.16 Aloss sapticisants 19 0.14 0.157 Coloroscombras charguar 11 0.07 0.081 0.081 Aloss sapticisants 19 0.14 0.139 Coloroscombras charguar 10 0.07 0.021 Icabarchity spitteria 18 0.13 0.131 0.132 0.621 Coloroscombras charguar 10 0.07 0.041 0.22 0.166 0.131 1.18 Cohooscombras charguar 10 0.070										
Erogus crossolar 25 0.17 0.075 0.04 Uraphysic projes 28 0.20 0.33 Alow sapidisona 17 0.11 0.15 Preprins frienamina 26 0.19 0.03 Alow sapidisona 17 0.11 0.32 Operator strutumes 25 0.19 0.03 Cyrinar carpio 16 0.11 2.33 0.23 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.19 0.14 0.20 1.15 <i>Frequisa strutumisa</i> 19 0.14 0.20 0.18 1.16 0.12 0.15 Errapics crossolar 10 0.12 0.63 0.19 0.18 0.19 <										0.6
Anguila costrata 19 0.13 2.01 1.15 Perprise transmatus 26 0.19 0.203 Urophycis fordiana 17 0.11 0.18 0.06 0.015 0.15 0.16 Urophycis fordiana 17 0.11 0.58 0.29 Perrisa alepidons 21 0.15 0.16 Chroscombrac chysurus 13 0.09 0.327 0.16 Alosa aestrivalis 19 0.14 0.037 Chroscombrac chysurus 13 0.09 0.285 0.01 Alosa aestrivalis 19 0.14 0.295 Chroscombrac chysurus 13 0.06 0.025 0.15 Europas crosolitas 19 0.14 0.205 Chroscombrac chysurus 11 0.07 0.037 0.022 0.01 Archosargue probatocephalia 17 0.12 0.881 Prinks triacentitis 8 0.05 0.020 0.11 Lapdord rhomboide 13 0.09 0.045 Porticis stristas 8 0.05 0.										0.3
Alox supplicitions 17 0.11 0.14 0.06 Cohessis strummant 25 0.15 0.076 Cyprints carpio 16 0.11 2.793 1.39 Chiorsscombas chrysuns 21 0.15 0.190 Cyprints carpio 16 0.11 2.793 1.39 Chiorsscombas chrysuns 21 0.15 0.196 Consection nebulous 13 0.09 0.038 0.014 Alosa saptidissima 19 0.14 0.206 Dayoits subma 12 0.08 0.851 5.41 Citharichthys spligheres 18 0.13 0.118 Gobioso strumosas 11 0.07 0.027 Citharichthys spligheres 18 0.12 0.851 Parints lingholis 8 0.05 0.045 0.02 Uraphysic floridana 19 0.07 7.515 Parints alighiotis 8 0.05 0.045 0.02 Lepisosttus soria 8 0.66 0.025 Adrona hegesta 7 0.05 0.057										0,2
Unophysis floridana 17 0.11 0.382 0.29 Peprinas alepidonas 23 0.17 0.130 Cyrnais con nebuloasa 13 0.09 0.327 0.16 Alexa aserinalis 20 0.14 0.097 Choroscombras chrysunas 12 0.08 0.028 0.01 Alexa aserinalis 20 0.14 0.047 Alosa astrinatis 12 0.08 0.028 0.01 Alexa aserinatis 19 0.14 0.189 Choroscombras chrysunas 11 0.07 0.047 0.02 Culturinits 17 0.12 0.68 Cyrinus cargin 11 0.07 0.047 0.02 Culturinits 17 0.12 0.68 Pernitis incentinits 9 0.66 0.022 0.01 Archoangrup probatocephalus 17 0.12 0.483 Morren sacatiarts 8 0.05 0.03 Dazyatis salaria 8 0.66 0.223 Pernitis incentinits inbulas 8 0.05 0.03 P										0.1
Cprimas carpio 6 0.11 2.793 1.39 Chorascombras chrysurus 21 0.15 0.16 Consolon methodosa 13 0.09 0.028 0.01 Aloar aerivalis 20 0.14 0.149 Choroscombras chrysurus 13 0.09 0.028 0.015 Erropus crossotus 19 0.14 0.149 Aloar aerivalis 12 0.08 0.851 5.41 Citharchitylis splopterus 18 0.13 0.118 Gobiess strumosus 11 0.07 0.022 0.01 Archoargys probatocephalas 17 0.12 0.435 Prinotons critoluis 11 0.07 0.022 0.01 Archoargys probatocephalas 16 0.12 0.435 Moren escatulis 8 0.06 0.220 0.11 Lagodon rhomboides 13 0.09 0.435 Astroscopus - graecum 7 0.05 0.057 0.03 Pomatomus salatics 8 0.06 0.215 Astroscopus - graecum 7 0.0										0.0
Cynascion nebulosis 13 0.09 0.327 0.16 Alexa astriadis 20 0.14 0.039 Aloxa astriadis 12 0.08 0.295 0.15 Erropss crossoitas 19 0.14 0.206 Dayatis subina 12 0.08 0.295 0.15 Erropss crossoitas 19 0.14 0.206 Dayatis subina 12 0.08 0.295 0.15 Erropss crossoitas 18 0.13 0.11 Chroscontinus 10 0.07 0.045 0.02 Uraphysis fordiana 16 0.12 0.452 Pontionis sultative 9 0.06 0.045 0.02 Uraphysis fordiana 16 0.12 0.452 Pontionis sultative 9 0.06 0.020 0.11 Lagodon rhombiolise 13 0.09 0.452 Partini dipidotas 8 0.05 0.03 Prinontias sultative assents 8 0.06 0.225 Ancoa hepsetus 7 0.05 0.05 0.03										0.0
Chlorascenhras chrysuns 13 0.09 0.028 0.01 Atoas aspidassma 19 0.14 0.149 Davyatis sabina 12 0.08 0.295 0.15 Eropas crossotis 19 0.14 0.296 Davyatis sabina 12 0.08 0.81 5.41 Ciharchitys spilopteras 18 0.13 0.118 Gobisos strumosas 11 0.07 0.02 Ichianra furcatus 17 0.12 0.63 Printo legitolus 9 0.06 0.200 11 Lagodor frombolides 13 0.09 0.495 Partino legitolus 8 0.05 0.001 Daystis sabina 9 0.07 7.515 Partino legitolus 8 0.05 0.03 Pernotaris valatarix 8 0.06 0.225 Astroscopus - graecum 7 0.05 0.05 0.03 Pernotaris valata 8 0.06 0.225 Astroscopus - graecum 7 0.05 0.045 0.02 Daystis valata 8									0.196	0.1
Alexa estivulis 12 0.08 0.295 0.15 Erropus crossolus 19 0.14 0.206 Dayatia subhina 12 0.08 0.81 5.41 Cithanichthys splopterus 18 0.13 0.14 0.26 Gobiesax strumous 11 0.07 0.047 0.02 101 Archoargues probability 17 0.12 0.82 Pernits tribulas 11 0.07 0.022 0.01 Archoargues fordana 16 0.12 0.432 Pernits tribulas 8 0.05 0.070 0.03 Dissibility 9 0.06 8.025 Parlits abgridotus 8 0.05 0.055 0.03 Perinotins sultatrix 8 0.06 0.225 Archoa hepsetus 7 0.05 0.055 0.03 Perinotins sultatrix 8 0.06 0.046 Cardorn rhomboides 6 0.044 0.022 Arcylopetia quadrocellata 6 0.044 0.032 Lagodon rhomboides 6 0.044									0.057	0.0
Dayati sabina 12 0.08 10.81 5.41 Cubinativitys galopterus 18 0.13 0.18 Gobiesox strumous 11 0.07 0.022 0.01 Archoargus probatocephilis 17 0.12 0.63 Fronous tribulas 11 0.07 0.022 0.01 Archoargus probatocephilis 17 0.12 0.63 Peprilus diephotus 9 0.06 0.022 0.01 Archoargus probatocephilis 10 0.435 Morone sacatilis 8 0.05 0.045 Doze Lepsotate soasus 8 0.06 0.234 Astroscopus y graecum 7 0.05 0.057 0.03 Pomatomus sultarix 8 0.06 0.011 Approbleminis hentzi 6 0.044 0.02 Arcylospetra quadrocellatiz 6 0.044 0.02 0.03 0.037	and the second					Alosa sapidissima	19	0.14	0.149	0.0
Gobesx stramosas 1 0.07 0.047 0.02 Lendmanno gradus 17 0.12 0.632 Prinotus tribulas 11 0.07 0.022 0.01 Archoargus probatosphalus 17 0.12 0.632 Prinotus tribulas 9 0.06 0.045 0.02 Urophysics floridana 16 0.12 0.432 Pomtomus saliatrix 9 0.06 0.020 0.01 Lagodon rhomboides 13 0.09 0.495 Anchoa hepsetus 8 0.05 0.05 0.03 Pomtomus saliatrix 8 0.06 0.285 Anchoa hepsetus 7 0.05 0.055 0.03 Prinotus tribulus 8 0.06 0.285 Anchoa hepsetus 7 0.05 0.055 0.03 Prinotus tribulus 8 0.06 0.285 Anchoa hepsetus 5 0.03 0.07 7 0.05 0.057 Lagodon rhomboide 6 0.04 0.228 0.11 Ancivospetia qancocelaia <t< td=""><td></td><td></td><td></td><td></td><td></td><td>Etropus crossotus</td><td>19</td><td>0.14</td><td>0.206</td><td>0.1</td></t<>						Etropus crossotus	19	0.14	0.206	0.1
Prinonus tribulas 1 0.07 0.022 0.01 Archosargus probaticocephalus 1 0.12 0.381 Peprilus triacanthus 9 0.06 0.045 0.02 Urophycis fordiana 16 0.12 0.432 Peprilus delpoidon 8 0.05 0.045 0.02 Lepisotens social 9 0.07 7.515 Peprilus delpoidons 8 0.05 0.045 0.02 Lepisotes asseus 8 0.06 0.285 Astroscopus systaccum 7 0.05 0.055 0.03 Pomatomus saltatrix 8 0.06 0.285 Astroscopus systaccum 7 0.05 0.055 0.03 Prinotus staltatrix 8 0.06 0.285 Astroscopus systaccum 7 0.05 0.055 0.02 Astroscopus systas 7 0.05 0.057 Lagdon rhombides 6 0.04 0.22 Astroscopus systas 6 0.04 0.227 Ancylopstita quadrocellata 4 0.03 0.037					5.41	Citharichthys spilopterus	18	0.13	0.118	0.0
Perplica triacanthas 9 0.66 0.035 0.72 Urbandy as products of private 1 0.12 3.801 Pomatomus saltarix 9 0.06 0.220 0.11 Lagodon rhomboides 13 0.09 0.405 Morone sexulis 8 0.05 0.070 0.03 Degissi subina 9 0.06 8.075 Astroscops yergreeum 7 0.05 0.067 0.03 Pomotus tribulus 8 0.06 0.025 Anchoa hepsetis 7 0.05 0.055 0.03 Prionotus tribulus 8 0.06 0.011 Hypoblennishentri 6 0.04 0.228 0.11 Anchoa hepsetis 7 0.05 0.046 Centropristis striata 6 0.04 0.023 Ancylopseti quadrocellata 6 0.044 0.028 Legioant rhomboides 5 0.03 0.171 0.36 Clainus puncturatus 6 0.044 0.032 Legioanta padarocellata 4 0.03 0.072 <t< td=""><td></td><td></td><td></td><td>0.047</td><td>0.02</td><td>Ictalurus furcatus</td><td>17</td><td>0.12</td><td>0.632</td><td>0.3</td></t<>				0.047	0.02	Ictalurus furcatus	17	0.12	0.632	0.3
Pomtomus saltatrix 9 0.6 0.20 0.11 Lagodon rhomboides 13 0.12 0.432 Morone saxatilis 8 0.05 0.045 0.02 Lepisoteus oscut 8 0.06 0.073 7.515 Peprilia delpidotas 8 0.05 0.045 0.02 Lepisoteus oscut 8 0.066 0.073 7.515 Astroscopus y-graecum 7 0.05 0.067 0.03 Priontous tribulus 8 0.066 0.025 Carnorritis striat 6 0.04 0.045 0.02 Selene vomer 7 0.05 0.057 Lagodon rhomboides 6 0.04 0.042 Ancylopsetia quadrocellata 6 0.044 0.032 Lagodon rhomboides 5 0.03 0.171 0.36 Intrans punctatus 6 0.044 0.032 Lagodon rhomboides 10.03 0.047 0.071 Centropristis philodelphica 4 0.03 0.014 Dorssoma cepepedianum 4 0.03 <			0.07	0.022	0.01	Archosargus probatocephalus	17	0.12	3.861	2.2
Morone sexatilis 8 0.05 0.070 0.033 Degvain subina 10 0.007 7.515 Peprilus alepidotus 8 0.05 0.043 0.02 Lepisostess scess 8 0.066 0.285 Acrisosopis sygreecum 7 0.05 0.057 0.03 Prinatorus saliatrix 8 0.06 0.285 Acricoa hepsetus 7 0.05 0.055 0.03 Prinontus tribulus 8 0.06 0.0285 Acricoa hepsetus 7 0.05 0.057 0.05 0.057 Legodon rhomboides 6 0.04 0.228 0.11 Ancioa hepsetus 7 0.05 0.057 Legodon rhomboides 6 0.04 0.029 Anciyopsetia quadrocellata 6 0.04 0.029 Keigli cephalus 5 0.03 0.177 0.36 Iclaturus purcetus 6 0.04 0.027 Dorosoma cepedianum 4 0.03 0.143 0.02 Cararax hippos 4 0.03			0.06	0.045	0.02	Urophycis floridana	16	0.12	0.432	0.2
Morone sexatilis 8 0.05 0.070 0.03 Dayatis salina 9 0.07 7,515 Perphis delpidotas 8 0.05 0.045 0.02 Lepisosteus osseus 8 0.06 0.235 Astroscopus y-graecum 7 0.05 0.067 0.03 Prinontus tribulas 8 0.06 0.235 Ancha hepsetus 7 0.05 0.064 0.22 Selene vomer 7 0.05 0.066 Contropristis striata 6 0.04 0.228 0.11 Anchoa hepsetus 7 0.05 0.057 Lagodon rhomboides 6 0.04 0.228 0.11 Anchoa hepsetus 6 0.04 0.028 Mugil cephalus 5 0.03 0.171 0.35 Ictaturus purcitas 6 0.04 0.027 Ancylopsetta quadrocellaa 4 0.03 0.037 0.02 Opishonema oglinum 5 0.04 0.014 Dorsoma cepedianum 4 0.03 0.016 Selene v			0.06	0.220	0.11	Lagodon rhomboides	13	0.09	0.405	0.2
Perfils slepidotus 8 0.05 0.045 0.02 Lepisosteu sossus 8 0.06 8.075 Astroscopus y spraceum 7 0.05 0.055 0.03 Pornatomus saltatrix 8 0.06 0.011 Hypsoblennius hentzi 6 0.04 0.045 0.02 Selene vomer 7 0.05 0.057 Lagodon rhomboides 6 0.044 0.02 Anchoa hepsetus 7 0.05 0.057 Lagodon rhomboides 6 0.044 0.02 Ancylopsetus quadrocellata 6 0.04 0.382 Meijl cephalus 5 0.03 0.717 0.36 Ictaturus punctatus 6 0.04 0.022 Ancylopsetta quadrocellata 4 0.03 0.037 Opistionema oglinum 5 0.04 0.014 Dorssona cepedianum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.166 Lepisoteus ossus 3 0.02 0.013 0.01 Lutyanus grisus <td>Morone saxatilis</td> <td>8</td> <td>0.05</td> <td>0.070</td> <td>0.03</td> <td>Dasyatis sabina</td> <td>9</td> <td>0.07</td> <td></td> <td>4.3</td>	Morone saxatilis	8	0.05	0.070	0.03	Dasyatis sabina	9	0.07		4.3
Astroscopus yegraecum 7 0.05 0.07 0.03 Pomatomus salutitic 8 0.06 0.285 Anchoa hegsetus 7 0.05 0.055 0.03 Prinontus tribulus 8 0.06 0.011 Hypsoblennius hentzi 6 0.04 0.022 Selene vomer 7 0.05 0.046 Centropristis striata 6 0.04 0.228 0.11 Anchoa hegsetus 7 0.05 0.037 Lagdon rhomboides 6 0.04 0.028 0.007 Outs stribulus 6 0.04 0.0382 Italiants platycephalus 5 0.03 0.179 0.16 Prionotus evolans 6 0.04 0.028 Ancylopsetta quadrocellata 4 0.03 0.33 0.02 Opishlon marginatum 5 0.04 0.014 Dorssoma cepedianum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.0166 Selene vomer 3 0.02 0.013 Out Lutymas grissus 4 0.03 0.024 0.054 S	Peprilus alepidotus	8	0.05	0.045	0.02	Lepisosteus osseus	8	0.06		4.7
Anchoa hepsetus 7 0.05 0.055 0.03 Prionotinus tribulus 8 0.06 0.011 Hypsohlennius hentri 6 0.04 0.042 Selene vomer 7 0.05 0.057 Lagodon rhomboides 6 0.04 0.228 0.11 Anchoa hepsetus 7 0.05 0.057 Lagodon rhomboides 6 0.04 0.022 Ancylopsetu quadrocellata 6 0.04 0.382 Meil cephalus 5 0.03 0.717 0.36 Ictalurus punctus 6 0.04 0.027 Ancylopsetu quadrocellata 4 0.03 0.037 0.02 Opisthonema oglinum 5 0.04 0.014 Dorssona cepedianum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.172 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.166 Selene vomer 3 0.02 0.01 Morone savatilitis 3 0.02 0.054 Lecinostomus argeneteus 2 0.01 <td>Astroscopus y-graecum</td> <td>7</td> <td>0.05</td> <td>0.067</td> <td>0.03</td> <td></td> <td>8</td> <td></td> <td></td> <td>0.1</td>	Astroscopus y-graecum	7	0.05	0.067	0.03		8			0.1
Hypsoblennius hentzi 6 0.04 0.045 0.02 Selene vomer 7 0.05 0.046 Centropristis striata 6 0.04 0.228 0.11 Anchoa hepsetus 7 0.05 0.057 Lagodon rhomboides 6 0.04 0.228 0.11 Ancylopsetta quadrocellata 6 0.04 0.032 Liadiurs platcatus 5 0.03 0.199 0.10 Prionottas evolans 6 0.04 0.027 Ancylopsetta quadrocellata 4 0.03 0.037 0.02 Opisthonema oglinum 5 0.04 0.014 Dorosoma cepedianum 4 0.03 0.143 0.062 Caranch hippos 4 0.03 0.166 Selene vomer 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.166 Selene vomer 3 0.02 0.013 Morone sazatilis 3 0.02 0.413 Chhardhtys splopterus 2 0.01 0.005 0.01	Anchoa hepsetus	7	0.05	0.055	0.03	Prionotus tribulus	8			0.0
Centropristis striata 6 0.04 0.228 0.11 Anchoa hepsetus 7 0.05 0.057 Lagadon rhomboides 6 0.04 0.044 0.02 Ancylopestia quadrocellata 6 0.044 0.0382 Migil cephalus 5 0.03 0.171 0.36 Ictalurus platycephalus 6 0.044 0.029 Ancylopestia quadrocellata 4 0.03 0.037 0.02 Opisthonema oglinum 5 0.044 0.014 Dorosoma cepedianum 4 0.03 0.149 0.07 Centropristis philodelphica 4 0.03 0.0142 Lepisosteus osseus 3 0.02 0.013 0.01 Lutjanus griseus 4 0.03 0.064 Selene vomer 3 0.02 0.013 0.01 Moros escarilis 3 0.02 0.143 Citharichthys splioptenas 2 0.01 0.005 <.0.01	Hypsoblennius hentzi	6	0.04	0.045	0.02	Selene vomer				0.03
Lagdon rhomboides 6 0.04 0.04 0.02 Ancylopsetta quadrocellata 6 0.04 0.0382 Mugil cephalus 5 0.03 0.199 0.10 Prinortus evolans 6 0.04 0.027 Icitalurus platycephalus 5 0.03 0.177 0.36 Icitalurus planctatus 6 0.04 0.227 Ancylopsetta quadrocellata 4 0.03 0.037 0.02 Opisthonema oglinum 5 0.04 0.014 Dorosoma cepedianum 4 0.03 1.243 0.62 Carax hippos 4 0.03 0.172 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.166 Selene vomer 3 0.02 0.01 Moros saxatilis 3 0.02 0.014 Gobionellus hastatus 2 0.01 0.005<	Centropristis striata	6	0.04	0.228	0.11					0.0
Muzil cephalus 5 0.03 0.199 0.10 Prionotus evolans 6 0.04 0.099 Ancylopsetta quadrocellata 4 0.03 0.037 0.02 Opisthomean oglinum 5 0.04 0.014 Dorosoma cepedianum 4 0.03 1.243 0.62 Caranx hippos 4 0.03 0.038 Ophidion marginatum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.166 Selene vomer 3 0.02 0.213 2.60 Ophidion marginatum 4 0.03 0.061 Membras marinica 2 0.01 0.021 0.01 Morone saxatilis 3 0.02 0.143 Citharichthys spilopterus 2 0.01 0.021 0.01 Dorosoma cepedianum 3 0.02 0.418 Gobionellis hastatus 2 0.01 0.022 0.01 Arrosoma cepedianum 3 0.02 0.418 Gobionellis hastatus 2 0.01 0.00	Lagodon rhomboides	6	0.04	0.044	0.02					0.22
Ictaluus platycephalus 5 0.03 0.717 0.36 Ictalurus punctatus 6 0.04 0.027 Ancylopsetta quadrocellata 4 0.03 0.037 0.02 Opishonema oglinum 5 0.04 0.014 Dorosoma cepedianum 4 0.03 0.143 0.62 Caranx hippos 4 0.03 0.0172 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.041 Membras martinica 2 0.01 0.005 <<0.01		5	0.03	0.199	0.10					
Ancylopsetta quadrocellata 4 0.03 0.037 0.02 Opisthonema oglinum 5 0.04 0.014 Dorosoma cepedianum 4 0.03 1.243 0.62 Caranx hippos 4 0.03 0.172 Depisoteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.041 Membras martinica 2 0.01 0.005 <0.01	Ictalurus platycephalus	5	0.03							0.01
Dorosoma cepedianum 4 0.03 1.243 0.62 Carax hippos 4 0.03 0.044 Ophidion marginatum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.014 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.016 Selene vomer 3 0.02 0.013 0.01 Lutjanus griseus 4 0.03 0.041 Membras martinica 2 0.01 0.005 <0.01		4	0.03							0.13
Ophidion marginatum 4 0.03 0.149 0.07 Centropristis philadelphica 4 0.03 0.018 Lepisosteus osseus 3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.017 Selene vomer 3 0.02 0.013 0.01 Lutjanus griseus 4 0.03 0.041 Membras martinica 2 0.01 0.02 0.01 Moroe suxailis 3 0.02 0.054 Eucinostomus argenteus 2 0.01 0.021 0.01 Dorosoma cepedianum 3 0.02 0.448 Gobionellus hastatus 2 0.01 0.005 < 0.01	Dorosoma cepedianum	4								0.1
Lepisosteus osseus3 0.02 5.213 2.60 Ophidion marginatum 4 0.03 0.116 Selene vomer3 0.02 0.013 0.01 Lutjanus griseus 4 0.03 0.041 Membras martinica2 0.01 0.005 <0.01 Morone saxatilis 3 0.02 0.054 Eucinostomus argenteus2 0.01 0.005 <0.01 Morone saxatilis 3 0.02 0.041 Gobionellus hastatus2 0.01 0.005 <0.01 Arius felis 3 0.02 0.042 Hysoblennius ionthas2 0.01 0.005 <0.01 Arius felis 3 0.02 0.042 Chaetodipterus faber1 0.01 0.223 0.11 Scomberomorus maculatus 2 0.01 0.004 Prionotus scitulus1 0.01 0.002 <0.01 Eleotris pisonis 2 0.01 0.024 Archosargus probatocephalus1 0.01 0.002 <0.01 Eleotris pisonis 2 0.01 0.014 Centropristis philadelphica1 0.01 0.007 <0.01 Chaetodipterus faber 2 0.01 0.016 Acipenser oxyrhynchus1 0.01 0.007 <0.01 Chaetodipterus faber 2 0.01 0.014 Acipenser oxyrhynchus1 0.01 0.007 <0.01 Chaetodipterus faber 2 0.01 0.016 Gobionellus shufeldi1 0.01 0.001										< 0.01
Selene vomer 3 0.02 0.013 0.01 Lutianus gresus 4 0.03 0.064 Membras martinica 2 0.01 0.005 <0.01										0.10
Membras martínica 2 0.01 0.005 <0.01										0.10
Eucinostomus argenteus 2 0.01 0.021 0.01 Dorosoma cepedianum 3 0.02 0.054 Citharichthys spilopterus 2 0.01 0.005 <0.01										0.02
Citharichthys spilopterus 2 0.01 0.005 <0.01 Arius felis 3 0.02 0.143 Gobionellus hastatus 2 0.01 0.093 0.05 Eucinostomus sp. 3 0.02 0.418 Hypsoblennius ionthas 2 0.01 0.007 <0.01									0.054	0.03
Gobionellus hastatus 2 0.01 0.093 0.05 Eucinostomus sp. 3 0.02 0.418 Hypsoblennius ionthas 2 0.01 0.007 <0.01									0.143	0.08
Hypsoblennius ionthas 2 0.01 0.007 < 0.01								0.02	0.418	0.24
Chaetodipterus faber 1 0.01 0.223 0.01 Prinontus scitulus 3 0.02 0.007 Prionotus scitulus 1 0.01 0.223 0.11 Scomberomorus maculatus 2 0.01 0.043 Archosargus probatocephalus 1 0.01 0.022 <0.01								0.02	0.042	0.02
Prionotus scitulus 1 0.01 0.002 < 0.01		1						0.02	0.007	< 0.01
Archosargus probatocephalus 1 0.01 0.338 0.17 Astroscopus y-graecum 2 0.01 0.024 Morone americana 1 0.01 0.338 0.17 Astroscopus y-graecum 2 0.01 0.011 Morone americana 1 0.01 0.484 0.24 Bagre marinus 2 0.01 0.011 Bagre marinus 1 0.01 0.144 0.07 Orthopristis chrysoptera 2 0.01 0.112 Bagre marinus 1 0.01 0.007 <0.01		1						0.01	0.043	0.03
Morone americana 1 0.01 0.484 0.24 Bagre marinus 2 0.01 0.011 Centropristis philadelphica 1 0.01 0.484 0.24 Bagre marinus 2 0.01 0.158 Bagre marinus 1 0.01 0.144 0.07 Orthopristis chrysoptera 2 0.01 0.158 Bagre marinus 1 0.01 0.007 <0.01		1					2	0.01	0.024	0.01
Motione unificand 1 0.01 0.484 0.24 Bagre marinus 2 0.01 0.158 Centropristis philadelphica 1 0.01 0.144 0.07 Orthopristis chrysoptera 2 0.01 0.112 Bagre marinus 1 0.01 0.007 <0.01		1				Astroscopus y-graecum	2	0.01	0.011	0.01
Centropistis philadelphica 1 0.01 0.144 0.07 Orthopistis chrysoptera 2 0.01 0.112 Bagre marinus 1 0.01 0.007 <0.01 Chaetodipterus faber 2 0.01 0.112 Acipenser oxyrhynchus 1 0.01 0.300 0.15 Mugil cephalus 1 0.01 0.100 Prionotus sp. 1 0.01 0.001 <0.01 Sciaenops ocellata 1 0.01 0.002 Gobionellus shufeldti 1 0.01 0.001 <0.01 Larimus fasciatus 1 0.01 0.002 Eucinostomus sp. 1 0.01 0.002 <0.01 Eucinostomus argenteus 1 0.01 0.002 Scophthalmus aquosus 1 0.01 0.002 <0.01 Morone americana 1 0.01 0.002 Perca flavescens 1 0.01 0.002 <0.01 Morone americana 1 0.01 0.008		1					2 .	0.01	0.158	0.09
Dagre maining 1 0.01 0.007 <0.01		1				Orthopristis chrysoptera	2	0.01		0.07
Actigative oxympticities 1 0.01 0.300 0.15 Mugil cephalus 1 0.01 0.100 Prionotus sp. 1 0.01 0.001 <0.01		1				Chaetodipterus faber	2	0.01		0.10
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		1					1			0.06
Gobionellus shufeldii 1 0.01 0.001 < 0.01 Larimus fasciatus 1 0.01 0.001 < Eucinostomus sp. 1 0.01 0.008 < 0.01		1				Sciaenops ocellata	1			< 0.00
Eucinostomus sp. 1 0.01 0.008 <0.01		1					1			< 0.01
Scophhalmus aquosus 1 0.01 0.002 < 0.01 Morone americana 1 0.01 0.338 Perca flavescens 1 0.01 0.017 0.01 Syngnathus louisianae 1 0.01 0.338 Chilomycterus schoepfi 1 0.01 0.002 < 0.01		1		0.008	< 0.01		1			< 0.01
Perca flavescens 1 0.01 0.017 0.01 Syngnathus louisianae 1 0.01 0.008 Chilomycterus schoepfi 1 0.01 0.002 <0.01		1	0.01	0.002	< 0.01		1			
Chilomycterus schoepfi 1 0.01 0.002 <0.01 Menidia menidia 1 0.01 0.002 Total 14,929 200.403 Membras martinica 1 0.01 0.004 Strongylura marina 1 0.01 0.004		1	0.01	0.017	0.01		1			0.20
Total 14,929 200.403 Membras martínica 1 0.01 0.002 < Strongylura marina 1 0.01 0.004 1 0.01 0.004 1 0.01 0.004 1 0.01 0.004 Trachinotus falcatus 1 0.01 0.004 0.01 0.004 0.01 0.004 <t< td=""><td>Chilomycterus schoepfi</td><td>1</td><td>0.01</td><td>0.002</td><td>< 0.01</td><td></td><td>1</td><td></td><td></td><td>< 0.01</td></t<>	Chilomycterus schoepfi	1	0.01	0.002	< 0.01		1			< 0.01
Strongylura marina 1 0.01 0.004 < Strongylura marina 1 0.01 0.075 Trachinotus falcatus 1 0.01 0.004 Trachinotus carolinus 1 0.01 0.004	Total	14,929					1			< 0.01
Trachinotus falcatus 1 0.01 0.004 Trachinotus carolinus 1 0.01 0.005				200,403			1			< 0.01
Trachinotus carolinus 1 0.01 0.005 <							1			0.04
							1			< 0.01
UTODAVCIS PDPID 1 0.01						Urophycis earlli	1	0.01	0.005	< 0.01

Urophycis earlli

Brevoortia smithi

1

1

0.01

0.01

0.001

0.268

< 0.01

0.16

	Total	Percent of	Total	Percent
Species	no.	total catch	weight	weight
South Santee RiverCont.				
Anguilla rostrata	1	0.01	0.150	0.09
Mugil curema	1	0.01	0.040	0.02
Gobionellus hastatus	1	0.01	0.009	0.01
Prionotus carolinus	1	0.01	0.001	< 0.01
Prionotus sp.	1	0.01	0.002	< 0.01
Cyprinus carpio	1	0.01	1.562	0.91
Gobionellus shufeldti	1	0.01	0.002	< 0.01
Diapterus olisthostomus	1	0.01	0.022	0.01
Chilomycterus schoepfi	1	0.01	0.085	0.05
Monacanthus hispidus	1	0.01	0.003	< 0.01
Total	13,826		171.372	

The total number of species of fishes and decapods varied over the 2-yr sampling period with the greatest number occurring in summer in both rivers (Fig. 2). Fewest species were collected during spring of 1975 when freshwater input and riverflow drastically increased. The dramatic drop in number of species was most noticeable at stations located upriver (NS07, NS11, and SS07). The total number of species captured was much lower during this time period than in spring of 1976 when no freshet occurred. During the 2-yr sampling period, more species were collected at stations nearest the mouths of both rivers.

The number of individual fish and decapod crustaceans, expressed in logarithms, showed patterns similar to the number of species when plotted over time (Fig. 2). In both rivers, numbers of individuals were greater during 1975, with peaks occurring in summer.

The number of species and number of individuals were compared to environmental factors such as bottom temperature, salinity, oxygen, turbidity, and depth using Pearson's productmoment correlation coefficient (Table 3). Based on these analyses, we found the number of species in the North Santee River to be significantly associated with bottom temperature and salinity in 1975 and with salinity and depth in 1976. In the South Santee River, the number of species was significantly associated with salinity during both years.

In the North Santee system, the number of individuals was positively correlated with bottom temperature in 1975, but there were no significant associations detected between number of individuals and environmental factors in 1976 (Table 3). The number of individuals captured in the South Santee system was positively correlated with bottom temperature and negatively correlated with oxygen in 1975 and 1976. A positive correlation with depth was found also in 1976.

Normal cluster analysis revealed that no strong differentiation of collections existed by river mile. Rather, collections made in the limnetic-euhaline zone were grouped with those from the limnetic-oligohaline zone indicating little stratification of the fauna according to salinity regime. In addition, an examination of the allocation of collections according to station and month indicated that association of the collections was not related to time of year. Based on similarity of faunal composition, we discerned three primary station groups by cluster analysis of data from the North Santee River: 1) a group in which collections at station NS01, NS04, and NS07 were represented by nearly equal numbers of collections; 2) a group in which collections at station NS01 predominated; and 3) a group which was most distinct from the other groups and was dominated by collections made at station NS11. Two major groupings of stations were indicated by cluster analysis for the South Santee River: 1) a group consisting mostly of collections from stations SS01 and SS04, and 2) a group consisting predominantly of collections from stations SS07 and SS11.

Species	Total no.	Percent of total catch	Total weight	Percent weight	Species	Total no.	Percent of total catch	Total weight	Percent weight
North Santee River:					South Santee River:				
Penaeus setiferus	34,998	90.08	121.703	48.38	Penaeus setiferus	10,431	78.80	44.333	44.98
Penaeus aztecus	1,556	4.00	13.393	5.32	Penaeus aztecus	1,726	13.04	10.242	10.39
Callinectes sapidus	1,318	3.39	114.709	45.60	Callinectes sapidus	568	4.29	42.763	43.39
Palaemonetes vulgaris	510	1.31	0.228	0.09	Macrobrachium ohione	120	0.91	0.414	0.42
Palaemonetes pugio	255	0.66	0.125	0.05	Palaemonetes pugio	90	0.68	0.047	0.05
Penaeus duorarum	95	0.24	1.036	0.41	Palaemonetes vulgaris	89	0.67	0.052	0.05
Panopeus herbstii	23	0.06	0.085	0.03	Trachypenaeus constrictus	83	0.63	0.071	0.07
Portunus gibbesii	19	0.05	0.016	0.01	Callinectes similis	31	0.23	0.184	0.19
Callinectes similis	18	0.05	0,161	0.06	Penaeus duorarum	17	0.13	0.077	0.08
Trachypenaeus constrictus	18	0.05	0.019	0.01	Clibanarius vittatus	16	0.12	0.024	0.02
Rhithropanopeus harrisii	17	0.04	0.013	0.01	Pagurus longicarpus	12	0.09	0.006	0.01
Clibanarius vittatus	13	0.03	0.024	0.01	Xiphopenaeus kroyeri	12	0.09	0.030	0.03
Portunus spinimanus	5	0.01	0.008	< 0.01	Rhithropanopeus harrisii	10	0.08	0.134	0.14
Macrobrachium ohione	3	0.01	0.028	0.01	Portunus spinimanus	7	0.05	0.104	0.11
Xiphopenaeus kroyeri	2	0.01	0.004	< 0.01	Callinectes similis or ornatus ¹	7	0.05	0.018	0.02
Panopeus occidentalis	2	0.01	0.002	< 0.01	Callinectes ornatus	5	0.04	0.031	0.03
Panopeus sp.	1	< 0.01	0.001	< 0.01	Panopeus herbstii	4	0.03	0.006	0.01
Alpheus heterochaelis	1	< 0.01	0.001	< 0.01	Portunus gibbesii	3	0.02	0.007	0.01
Total	20.054				Alpheus heterochaelis	2	0.02	0.002	< 0.01
Iotai	38,854		251.556		Acetes americanus	1	0.01	0.001	< 0.01
					Ovalipes ocellatus	1	0.01	0.002	< 0.01
					Eurypanopeus depressus	1	0.01	0.001	< 0.01
					Xanthidae ²	1	0.01	0.008	0.01
					Total	13,237		98.557	

Table 2.—Total number and total weight (kg) of decapod Crustacea collected from 1975 and 1976 in the North and South Santee Rivers. Species are listed in order of abundance and data are pooled over the 2-yr sampling period.

Field identification.

²Specimen damaged and unidentifiable, not included in analyses.



Figure 2.—Monthly fluctuations in number of species and number of individuals (log transformed) of fishes and decapod crustaceans at sampling sites in the North and South Santee Rivers, 1975-76.

Table 3.—Correlation between $\ln (n + 1)$ transformed values of number of species and number of individuals of fishes and decapods in relation to environmental factors. r = Pearson product-moment correlation coefficient; n = number of observations.

		19	75	1976				
	North San	South Santee		North Santee		South Santee		
Environmental factor	r	n	r	n	r	n	r	n
	Nur	nber	of species		Nur	nber	of species	
Bottom temperature (°C)	0.4270*	45	0.1646	36	-0.0225	48	-0.0769	48
Salinity (%)	0.3984*	45	0.4185*	36	0.5437*	48	0.5929*	48
Oxygen (mg/liter)	-0.1575	45	-0.0457	36	-0.0110	48	0.0074	48
Turbidity (FTU)**	0.2972	45	0.0849	36	-0.0550	48	-0.2213	48
Depth (m)	-0.3045	45	0.0170	36	-0.6029*	48	0.2420	48
	Numt	ber o	f individuals	5.	Numb	er of	individuals	
Bottom temperature (°C)	0.5003*	45	0.5158*	36	0.0239	48	0.3385*	48
Salinity (%)	0.2006	45	0.1605	36	0.0187	48	0.2766	48
Oxygen (mg/liter)	-0.0768	45	-0.3710*	36	-0.0614	48	-0.3089*	48
Turbidity (FTU)	0.1575	45	-0.2985	36	0.0142	48	-0.0146	48
Depth (m)	-0.1624	45	-0.1052	36	-0.1766	48	0.4652*	48

*Significant ($\rho \neq 0$) at $\alpha = 0.05$

**FTU = Formazin Turbidity Units.

Table 4.—Groups formed from cluster analysis of species of fishes and decapod Crustacea collected in the North and South Santee Rivers from 1975 and 1976. Dendrograms are not shown.

North Santee R.	South Santee R.
Group A	Group A
Lagodon rhomboides	Ophidion marginatum
Alpheus heterochaelis	Ancylopsetta quadrocellata
Eucinostomus argenteus	Urophycis regia
Group B	Urophycis floridana Etropus crossotus
Prionotus tribulus	Alosa sapidissima
Dasyatis sabina Callinectes similis	Cynoscion nebulosus
Gobiesox strumosus	Archosargus probatocephalus
Etropus crossotus	Lutjanus griseus
Hypsoblennius hentzi	Lagodon rhomboides
Xiphopenaeus kroyeri	Group B
Group C	Dasyatis sabina
Astroscopus y-graecum	Citharichthys spilopterus
Ancylopsetta quadrocellata	Caranx hippos
Ophidion marginatum	Eucinostomus sp.
Urophycis floridana	Centropristis philadelphica
Centropristis striata	Group C
Urophycis regia	Hypsoblennius ionthas
Penaeus duorarum	Trachypenaeus constrictus
Group D	Menticirrhus americanus
Arius felis	Clibanarius vittatus
Peprilus alepidotus	Hypsoblennius hentzi
Callinectes ornatus	Penaeus duorarum
Selene vomer	Centropristis striata
Portunus gibbesii	Opsanus tau
Pagurus longicarpus	Symphurus plagiusa
Clibanarius vittatus	Gobiesox strumosus
Opsanus tau	Paralichthys dentatus
Trachypenaeus constrictus	Group D
Menticirrhus americanus	Pomatomus saltatrix
Group E	Peprilus triacanthus
Alosa sapidissima	Anchoa hepsetus
Paralichthys dentatus	Chloroscombrus chrysurus
Cynoscion nebulosus	Panopeus herbstii
Mugil cephalus	Group E
Dorosoma petenense	Cynoscion regalis
Gobionellus hastatus	Penaeus aztecus
Rhithropanopeus harrisii	Stellifer lanceolatus
Peprilus triacanthus	Callinectes similis
Anchoa hepsetus	Peprilus alepidotus
Panopeus herbstii	Selene vomer
Pomatomus saltatrix	Lepisosteus osseus
Chloroscombrus chrysurus	Rhithropanopeus harrisii
Group F	Macrobrachium ohione
Lepisosteus osseus	Group F
Cyprinus carpio	Ictalurus punctatus
Palaemonetes pugio	Ictalurus furcatus
Alosa aestivalis	Alosa aestivalis
Ictalurus punctatus	Group G
Ictalurus furcatus	Anchoa mitchilli
Macrobrachium ohione	Bairdiella chrysoura
Anguilla rostrata	Callinectes sapidus
Morone saxatilis	Penaeus setiferus
Group G	Paralichthys lethostigma
Ictalurus catus	Brevoortia tyrannus
Trinectes maculatus	Leiostomus xanthurus
Micropogonias undulatus	Micropogonias undulatus
Anchoa mitchilli	Trinectes maculatus
Penaeus setiferus	Ictalurus catus
Bairdiella chrysoura	Palaemonetes vulgaris
Cynoscion regalis	Palaemonetes pugio
Stellifer lanceolatus	Dorosoma petenense
Penaeus aztecus	
Symphurus plagiusa	
Callinectes sapidus	
Paralichthys lethostigma	
Brevoortia tyrannus	
Leiostomus must	

Leiostomus xanthurus Palaemonetes vulgaris The classification based on the quantitative similarities of distribution of species found in the North and South Santee Rivers produced the species groups shown in Table 4. In order to determine affinity of species assemblages along the estuarine gradient, we compared species group constancy and fidelity among the eight stations occupied in the North and South Santee Rivers during 1975 and 1976 (Fig. 3). This was deemed preferable to comparing site groups determined by cluster analysis with species groups because site groups broadly overlapped and were not clearly separated by cluster analysis according to salinity regimes within the estuary.

In the North Santee River, one species group (G) was consistently encountered at stations NS01 and NS04, with slight decline in constancy at NS07 and NS11 (Fig. 3). Species in this group were not restricted in their distribution to any station location but were ubiquitous over the sites sampled, which is an indication of their apparent euryhalinity. The other species groups were not consistently collected at any of the stations, as indicated by low constancy. Species group B, which is largely composed of coastal marine fishes, was entirely restricted to station NS01, which suggests the stenohaline nature and transient occurrences of these fishes within the estuary. Other groups (A and D) were also apparently composed of marine species which were not able to penetrate far into the estuary. Group E species were associated with intermediate to higher salinities and did not



Figure 3.—Two-way coincidence tables of constancy and fidelity which compare species associations with stations in the North and South Santee Rivers, 1975-76. The species associations, designated alphabetically, resulted from cluster analysis of species (dendrogram not shown) collected from the Santee system. Species comprising these associations are listed in Table 5.

occur at stations upriver from NS04. Members of group F were not found downestuary of station NS07.

In the South Santee River, species in group G were considered to be ubiquitous over all sites. The constancy of these species ranged from high at stations SS04 and SS07 to moderate at SS01 and SS11; however, species in this group were not restricted to any station location. Groups A, C, and D included species which were associated with higher salinity areas in proximity to the river mouth. Stenohaline marine species in these groups included sheepshead, Archosargus probatocephalus; black sea bass, Centropristis striata, butterfish, Peprilus triacanthus; and Atlantic bumper, Chloroscombrus chrysurus. Group B species were not consistently collected at any station location and displayed low fidelity to stations SS04 and SS11. These species are generally considered to be marine in origin and their penetration as far as SS11 is unusual. Species which were associated with higher and intermediate salinities constituted group E. These were found at all stations except SS11, but were not consistently collected at any site. Group F contained the stenohaline freshwater species, Ictalurus punctatus and I. furcatus, and the anadromous species, blueback herring, Alosa aestivalis, which were restricted to station SS11.

Based on results of the two-way coincidence table (Fig. 3), it was possible to distinguish four assemblages of fishes and decapod Crustacea in both the North and South Santee Rivers. The first assemblage consisted of euryhaline species which occurred throughout both rivers and included the fishes Anchoa mitchilli; Brevoortia tyrannus; Trinectes maculatus; Micropogonias undulatus; Leiostomus xanthurus; Bairdiella chrysoura; southern flounder, Paralichthys lethostigma; and I. catus; and the decapods Penaeus setiferus, Callinectes sapidus, and Palaemonetes vulgaris, grass shrimp. Coastal marine species, which may penetrate into the estuary for short periods of time, constituted the second assemblage. Species in this category included the fishes Centropristis striata; skilletfish, Gobiesox strumosus; feather blenny, Hypsoblennius hentzi; southern kingfish, Menticirrhus americanus; and pinfish, Lagodon rhombiodes-and the decapods pink shrimp, Penaeus duorarum, and humpback shrimp, Trachypenaeus constrictus. The third assemblage consisted of species which can tolerate a range of intermediate to low salinities. Rhithropanopeus harrisii, mud crab, which occurs in both the North and South Santee Rivers, was distributed in this manner. The fourth assemblage was composed of the stenohaline freshwater species Ictalurus punctatus, I. furcatus, and the anadromous species, Alosa aestivalis.

Although the formation of these categories is based on distributional patterns formed from an actual collection of the organisms, it remains an artificial attempt at forcing species into designated groups based on their general affinities within the estuary. Therefore, it is possible and certainly probable that species will encounter others outside their group and may even form peripheral associations. This is especially true of the euryhaline species which are capable of widespread penetration of the estuarine environment.

Temporal and Spatial Distributions-Fishes

Temporal and spatial distributions for four abundant species of fishes—Micropogonias undulatus, Anchoa mitchilli, Ictalurus catus, Bairdiella chrysoura, and Trinectes maculatus are compared in Figures 4-6. A summary table of the distribution and lengths for all species collected is available from the authors.

Micropogonias undulatus, Atlantic croaker.-The Atlantic croaker was found throughout both rivers, although its presence at stations varied over the 2-yr sampling period (Fig. 4A). In the North Santee River, number and biomass of the Atlantic croaker were greatest during spring 1976 at NS07. This is in marked contrast to spring 1975 when none were collected at this station or at NS11 further upriver. The absence of croaker at these stations in 1975 may be attributed to the significant alteration of physicochemical properties by the freshet in spring 1975. A similar decrease in abundance was not noted in the South Santee River, but failure to occupy SS11 during 1975 precludes a true assessment of freshet effects far upriver. The apparent absence of Atlantic croaker at upriver stations in the North Santee River during fall 1976 cannot be readily attributed to any hydrographic parameter but may reflect a lag in recruitment of young fish during this period.

Length-frequency distributions (not shown) indicated that sizes of most Atlantic croaker available to our bottom trawls were <10 cm in both rivers during all seasons. The predominance of smaller fish accounts for the low biomass observed for Atlantic croaker. Young fish, 4-16 cm, were prevalent in both rivers during fall and winter. A few larger fish which ranged from 12 to 26 cm were also present, but their numbers were low, which could reflect gear avoidance, movement away from the channel, or emigration from the estuary. Size of young Atlantic croaker had increased to a mode of 8-9 cm by summer and abundance had increased. Others (Haven 1957; Hansen 1969; Hoese 1973; Shealy et al. 1974; Chao and Musick 1977) have noted that small Atlantic croakers are present in different estuarine systems along the east coast throughout much of the year. The abundance of young fish in the Santee system is probably related to the long spawning season of the Atlantic croaker (Chao and Musick 1977), which may be more protracted in South Carolina waters than in temperate northern estuaries, although our choice of sampling gear, biased toward capture of smaller fish, is undoubtedly also a factor.

Anchoa mitchilli, the bay anchovy.—Anchoa mitchilli was found at all stations in both the North and South Santee Rivers sometime during the 2-yr sampling period, but catches were generally greater in the South Santee River (Fig. 4). Abundance of *A. mitchilli* appeared to be lowest at low-salinity stations located further upriver in both rivers. This decreased abundance was especially noticeable in spring and summer. During these seasons, bay anchovy were found at more seaward stations within the estuary. This distributional pattern is similar to that observed in the Edisto and Cooper Rivers, S.C. (Shealy et al. 1974), and York River, Va. (Markle 1976).

Length-frequency distributions for *A. mitchilli* were strongly bimodal with smaller (20-35 mm) and larger (50-75 mm) fish cooccurring during most seasons (not shown). These data do not indicate an influx of small fish into the population during a particular season, such as summer (Hoese 1973; Shealy et al. 1974), but suggest that small fish are present in the Santee system throughout the year. Multiple spawns (Hoese 1965) or a protracted spawning season (Hildebrand and Cable 1930) best explain the bimodality of frequencies observed for bay anchovy in the Santee system. Similar findings were noted by Hoese (1965), who believed that *A. mitchilli* spawns during all seasons in Texas and probably is short-lived. In addition, Mansueti and



Figure 4.—Abundance of A) Micropogonias undulatus and B) Anchoa mitchilli by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total number of collections at a station.

Hardy (1967) found sexually mature individuals at 2.5 mo of age in the Chesapeake Bay system.

Ictalurus catus, white catfish.—The distribution of *I. catus* was obviously influenced by salinity since catches declined markedly at higher salinity stations (Fig. 5A). In the North Santee River, catches of *I. catus* were greatest during all seasons at stations furthest upriver. Distributional patterns in the South Santee River were similar in that *I. catus* seldom occurred at higher salinity stations. Shealy et al. (1974) found no *I. catus* at estuary mouths of the North and South Edisto or Charleston

Harbor-Cooper Rivers. The infrequent occurrence of white catfish at the mouth of the Santee River probably reflects the often low-salinity nature of the Santee River and the subsequent penetration by lower salinity species. High biomass of *I. catus* corresponded with peak numerical abundance in both rivers. Length-frequency distributions (not shown) showed that most white catfish collected in the Santee system were < 100 mm, although the length range extended from 10 to 370 mm. Based on an age-growth study of *I. catus* from South Carolina (Stevens 1959), the fishes < 100 mm are not older than 2 yr. In the North Santee River, young-of-the-year fish (< 50 mm) were



Figure 5.—Abundance of A) Ictalurus catus and B) Bairdiella chrysoura by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total number of collections at a station.

prevalent in summer, which is coincidental with the spawning period of *I. catus* in South Carolina (Stevens 1959).

Bairdiella chrysoura, the silver perch.—*Bairdiella chrysoura* was present in the Santee system during all seasons, although abundance tended to increase during fall and summer in the South Santee River (Fig. 5). Silver perch showed no apparent preference for a particular portion of the salinity regime in the middle and lower reaches of the estuary since they were collected

at all stations; however, catches did decline at the stations located further upriver (NS11 and SS11). *Bairdiella chrysoura* taken from the Santee system were young-of-the-year fish (Shealy et al. 1974; Chao and Musick 1977) within the size range of 20-100 mm.

Trinectes maculatus, **hogchoker**.—*Trinectes maculatus* was ubiquitous in the Santee system during all seasons (Fig. 6). Catches were greatest during fall in both the North and South Santee Rivers. Lower catches tended to be associated with sta-



Figure 6.—Abundance of *Trinectes maculatus* by station and season for the North and South Santee Rivers. Ratios over bars indicate number of collections where fishes were captured to total number of collections at a station.

tions in proximity to the river mouths, which suggests an avoidance of euhaline areas by this fish. Lengths of hogchokers ranged from 20 to 175 mm, but most individuals were < 80 mm. These specimens probably represent young-of-the-year fish (Dovel et al. 1969) which appear during all seasons due to the ex-

tended spawning season of this species in the Carolinas (Hildebrand and Cable 1938).

Temporal and Spatial Distributions-Decapods

Distributional patterns of the most abundant decapod crustaceans, *P. setiferus*, *P. aztecus*, and *Callinectes sapidus*, are shown in Figures 7 and 8.

Penaeus setiferus, white shrimp.-Catches of white shrimp were seasonal, with most individuals occurring in the Santee system during the fall and summer. Though common in both rivers, numerical abundance and biomass of white shrimp were greater in the South Santee River (Table 2), and catches appeared to be lower at the extreme upriver stations. This was particularly evident during winter and spring. Length-frequency distributions showed young-of-the-year white shrimp were present during summer in both rivers (Fig. 7). Sizes of shrimp collected increased during the other seasons, with the largest individuals collected in the spring. Similar findings were noted by Bishop and Shealy (1977) in a study of penaeid shrimp from South Carolina estuaries. They found that the largest numbers of shrimp were small, whereas larger individuals, which may be derived from the overwintering population or from an immigrating offshore population, occurred during fall and spring.

Penaeus aztecus, brown shrimp.-Brown shrimp were most abundant in spring and summer (Fig. 8). These brown shrimp were rare in winter trawl collections. In other South Carolina estuaries, Bishop and Shealy (1977) noted that catches of brown shrimp were strongly seasonal, with most individuals collected in summer. The absence of brown shrimp in trawl catches during the winter months does not indicate that they are absent entirely from the estuary. Postlarval shrimp first enter South Carolina estuaries in January and are most abundant in February and March (Bearden⁶). Because it appears that postlarval white shrimp, and perhaps also brown shrimp, primarily occupy the shallow edges and creeks of estuaries where cover and preferred food are available (Bishop and Shealy 1977), we may have failed to sample this component of the shrimp population by restricting our collecting to the channel. It is also probable that 9-12 mm postlarvae are not retained by our 6 m otter trawl. Examination of length-frequency distributions (not shown) for brown shrimp collected in the Santee system showed a total absence of postlarvae in our trawl collections. Shrimp ranged from 30 to 145 mm, with most individuals in the 55-90 mm size range. The abundance of brown shrimp was also related to station location and, hence, salinity as reflected in only one occurrence of P. aztecus at the extreme upriver sites.

Callinectes sapidus, **blue crab.**—The blue crab was caught throughout the North and South Santee Rivers during all seasons. Catches did not reflect strong seasonal changes, although fewer blue crabs were collected in summer in the North Santee River. Catches also appeared to be related to sampling location, with fewer blue crabs being caught at upriver stations. Size-frequency distribution of blue crabs covered a wide range of sizes from 15 to 195 mm, with smaller crabs (≤ 60 mm) occurring in fall.

^{*}Bearden, C. M. 1961. Notes on postlarvae of commercial shrimp (*Penaeus*) in South Carolina. Contrib. Bears Bluff Lab. No. 33, 8 p.





Biomass Estimates

The estimated biomass, expressed in kg/ha, for fishes from the North and South Santee Rivers was lower than biomass estimates reported for other estuarine systems along the Gulf and east coasts of the United States (Table 5). Greatest biomass was obtained for northern temperate estuaries such as Narragansett Bay, R.I. (Oviatt and Nixon 1973), and Mystic River, Mass. (Haedrich and Haedrich 1947), while the number of fish per hectare was comparable between these northern estuaries and those in South Carolina. The reliability of our biomass estimates is confirmed by the identical value (3.8 kg/ha) obtained by Shealy et al. (1974) in other South Carolina estuaries.

Table 5.—Estimates of density and number of individuals/hectare for fishes caught by trawls from estuaries along the Gulf and east coasts of the United States.

Geographic area	Biomass (kg/ha)	Density (no./ha)	Gear	Reference
Mystic River, Mass.	26.16	462	4.8 m semiballoon trawl	Haedrich and Haedrich (1974)
Narragansett Bay, R.I.	31.68	290	9.2 m balloon trawl	Oviatt and Nixon (1973)
North Santee River, S.C.	3.9	287	6 m semiballoon trawl	present study
South Santee River, S.C. Cooper River — Charleston	3.8	303	6 m semiballoon trawl	present study
Harbor and Edisto system, S.C.	3.8	433	6 m semiballoon trawl	Shealy et al. (1974)
Doboy Sound, Ga.	10.7	4,190	12.2 m balloon shrimp trawl	Hoese (1973)
Galveston Bay, Tex.	16.57	8,511	3 m otter trawl	Bechtel and Copeland (1970)



Figure 8.—Abundance of A) Penaeus aztecus and B) Callinectes sapidus. Ratios over the bars indicate number of collections where shrimp were captured to total number of collections at a station.

We obtained the following density estimates of decapod Crustacea from the Santee system:

	Biomass (kg/ha)	Density (no./ha)	
South Santee	5.5	836	
North Santee	1.9	255	
		000	

These estimates are comparable to 6.1 kg/ha and 1,190 individuals/ha reported by Hoese (1973) for all invertebrates collected from Doboy Sound, Ga.

DISCUSSION

The distributional patterns of estuarine fishes and decapod Crustacea are influenced by numerous environmental factors. Factors such as salinity (e.g., Gunter 1938, 1945, 1961; Kilby 1955; Kinne 1966; Copeland and Bechtel 1974), temperature (Gunter and Hildebrand 1951; Reid 1954; Kinne 1963), substrate and detritus (Carr and Adams 1973; Mills 1975; Livingston et al. 1977), and river discharge (Aleem 1972; Ruello 1973; Livingston et al. 1977; Glaister 1978) influence animal distributions, with the extent of these influences dependent on spatial (habitat) dimensions as well as individual and specific tolerances. Laboratory studies are generally concerned with the interrelationships between biological response and multiple environmental factors acting in concert. However, application of laboratory methodology to ecological field studies is often quite difficult (Alderdice 1972), especially when dealing with mobile organisms such as fishes and decapods. The interpretation of results concerning distribution of these organisms within the Santee system is no exception to this difficulty. Nevertheless, it is possible to make some interpretation of community stability and specific distributional patterns based on the information collected over the intensive 2-yr sampling period.

The freshet in spring of 1975 appeared to have the most pronounced singular effect on species composition and abundance. The total number of species collected was lower at that time than at any other during the sampling period. This was particularly noticeable at the upriver stations. Abundance of individual species such as Micropoganias undulatus, Anchoa mitchilli, and Trinectes maculatus were also lower at these stations, which also may be attributable to flood effects. The effects on the decapod crustaceans and other numerically dominant fishes were not obvious. Calder et al. (1977) also reported alterations of species composition and density among benthic organisms collected from the Santee system during 1975 and 1976; however, they noted that the flood most affected benthos in the lower, usually more saline reaches of the river where an increase in species normally associated with greater freshwater intrusion into the lower estuary occurred. Others (Andrews 1973; Boesch et al. 1976) have noted that effects of lowered salinity from floods are greatest among meso- or polyhaline species, but the magnitudes of the effects differ for epifaunal, infaunal, and highly motile organisms. The depressed species number observed by us may reflect the tendency of fishes and decapods to escape from areas whose salinity is drastically lowered by floodwaters or, in the case of juveniles and small-bodied species, may be attributed to their being flushed downstream and out of the system.

The positive correlation between salinity and number of species in the Santee system agrees with results obtained by Gunter (1961), who noted that the number of species increased toward the lower reaches of estuaries where there occurred a mixture of euryhaline and marine stenohaline species. Hoff and Ibara (1977) found that in a New England estuary the number of species was greatest at stations which had the greatest fluctuation in salinity. Both species number and the community assemblages defined by us for the Santee system reflect increased diversity with proximity to the river mouth. Also, most assemblages defined for the Santee system consisted partially of euryhaline species. This is not an unusual occurrence within estuaries. Pearse (1936) noted that the estuarine fauna consists of marine or marine-derived species, and Weinstein (1979) stressed how depauperate the shallow marsh estuarine fauna would be if all transient marine species were removed. The distribution of the endemic estuarine species appeared to be more restricted than that of the marine transients. Nevertheless, we observed no abrupt faunal changes along the salinity gradient in the Santee system. Rather, the faunal assemblages overlap and do not exist as sharply delineated groups. This no doubt results from the different tolerances of juveniles and adults; the effect of salinity, in concert with other factors, on reproduction; and the highly compressed nature of salinity regimes in South Carolina estuaries as compared with many estuaries elsewhere.

As indicated by Pearson product-moment correlations, temperature, depth, and dissolved oxygen also affect community composition and abundance. The positive correlation of number of species and number of individuals with temperature is not at all surprising when one considers that species composition was most diverse during summer in both rivers. The huge influx of Penaeus setiferus and, to a lesser extent, P. aztecus into the estuary during summer and fall probably accounts for this correlation. The association between depth and number of species and individuals is more difficult to explain. All collections were made in the channel where depths ranged from 2 to 8 m in the North Santee River and from 1 to 5 m in the South Santee River. Although the range in depths sampled is slight, sufficient salinity stratification may exist in the Santee system so that higher salinity water occurs in the deeper channel regions. This may indeed be true for the South Santee River which receives less freshwater input. Also, salinity stratification may be greater on the floodtide, where samples were collected (Mathews footnote 4.) Since there is a positive correlation of species number and abundance, the correlation between depth and species composition and abundance is most likely a secondary effect. Similarly, the negative association between dissolved oxygen and abundance which was noted only for the South Santee River may be explained by lower dissolved oxygen values in the deeper, more saline channel areas. We realize the correlations are simplistic and that misinterpretation can result from speculating about cause and effect relations in correlation analysis (Sokal and Rohlf 1969). We are merely presenting this information as untested hypotheses.

Peaks of abundance for the numerically dominant species were not generally consistent over the 2-yr sampling period, but peaks of maximum richness (number of species) consistently occurred in summer. This observation compares favorably with those of Livingston et al. (1976) for fishes and invertebrates of Apalachicola Bay, Fla. They noted a relatively stable appearance of organisms from year to year, but considerable within-species variability in annual abundance. Temporal partitioning by our dominant species was not as noticeable as that described by Livingston et al. (1976). Dominant fishes were present in the Santee system throughout the year and showed fairly equitable abundances, although M. undulatus and A. mitchilli dominated our catches in winter and spring. Penaeus setiferus was much more regular in its pattern of appearance and abundance. Although regular fluctuations in species composition over time may indicate that an estuary is not stressed (Livingston et al. 1976), we feel that the year-round presence of stress-tolerant estuarine species better indicates temporal stability than overall stability of the estuarine system. We relate such occurance to a protracted spawning season in warm temperate areas which enable some element of the population, probably juveniles, to be present in the estuary year-round.

As the length-frequency polygons for selected species showed, the Santee system fish fauna captured during this study are primarily composed of immature fishes. Some larger mature individuals were collected, but the Santee system functions strongly as a juvenile fish habitat. The importance of estuaries as nursery areas is well documented (Gunter 1961; Wallace and Van der Elst 1975; Livingston et al. 1976; Weinstein 1979), and the attraction of young fish to estuaries is attributed to physiological suitability in terms of physiochemical features, an abundance of food, and protection from predators (Gunter 1961; Van Engel and Joseph 1968⁷; Wallace and Van der Elst 1975).

Although Wallace and Van der Elst (1975) and Livingston et al. (1976) also found that juveniles predominated in their samples, we suspect that sampling design and gear selectivity may have biased our results toward juvenile fishes. Our choice of fixed stations is certainly biased and lends itself to sampling error that would have been eliminated or reduced by a stratified random design (Markle 1976). We are, therefore, not able to determine the influence of movements by the fauna between the shoals and the channel. Because trawling is inherently variable (Taylor 1953), a repetitive method of collection would have allowed for statistical analysis of sampling efficiency to determine whether hauls taken at different times in different places did indeed have significantly different catches (Barnes and Bagenal 1951; Livingston 1976). However, even with successive samples, it is difficult to determine whether variability arises from the spatial distribution of the organisms or from the gear utilized (Taylor 1953). The susceptibility of organisms to fishing gear undoubtedly has influenced perception of spatial and temporal patterns (Markle 1976). The relatively small, fine-mesh bottom trawl used in our study is selective toward capture of slower, smaller fish. The relative absence of great numbers of older, larger fish from our trawl catches cannot be attributed entirely to migration or habitat selection, but in all likelihood reflects at least partial avoidance or escapement from the 6 m trawl (Shealy et al. 1974).

Habitat differences between adult and juvenile fishes may also account for the lack of large fish in our samples. Habitat preference varies with the species and also with age (Wallace and Van der Elst 1975; White and Chittenden 1976), so that feeding and residential grounds of adult fishes often are separate from their spawning grounds and nurseries. If spatial separation exists in South Carolina waters, then our survey was biased toward collection of juveniles found primarily in the channel. However, tidal creeks of the Cooper River which are comparable in salinity to those near the intermediate and upriver stations occupied in the Santee system were dominated by young-of-theyear marine euryhaline species such as M. undulatus, A. mitchilli, L. xanthurus, B. chrysoura, and Paralichthys lethostigma (Turner and Johnson 1974). Although the importance of tidal creeks in the Santee system as nursery areas can only be inferred, it is likely that the limitation of our survey to the channel resulted in minimal estimates of juvenile abundance for the river system.

The lower estimated biomass of fishes in the Santee system and other South Carolina estuaries is a direct function of the predominance of juvenile fishes in our catches and the efficiency of the sampling gear used. Whereas the density of fish from this area compares favorably with other regions, the biomass is much less. The large biomass of fishes in New England estuaries is primarily due to large catches of winter flounder, *Pseudopleuronectes americanus* (Oviatt and Nixon 1973; Haedrich and Haedrich 1974). A comparison of biomass and density estimates from this study and others (see Table 5) which used small trawls towed in the channel with investigations which incorporated shallow tidal creek and marsh sampling (e.g., Turner and Johnson 1974) indicates that the most productive areas are the marsh-creek habitat. Because these areas of the system were not sampled and the efficiency of our gear was low, our biomass and density estimates should also be considered minimal.

The presence of juvenile fishes in the Santee system is especially important in considerations concerning the effects of rediversion. Juvenile stages of resident species and many estuarine transient species are tolerant of and may even be most abundant in lower salinity water (Gunter 1961). Therefore, we believe that the nursery habitat for resident estuarine fishes will not be detrimentally affected and may be increased by rediversion.

Because rediverted flow of water through the Santee system will be moderate compared with the tremendous discharge of freshwater (9,100 m³/s) put into the Chesapeake Bay estuarine system by Hurricane Agnes (Chesapeake Bay Research Council 1973), we do not anticipate that juvenile fishes will be passively swept from the Santee Rivers into the coastal area. In contrast, the food supply of fishes may be altered in that supplies of benthic organisms could increase in oligohaline and brackish water areas but decrease in lower reaches of the river. This effect could be particularly detrimental if it occurred during summer. Andrews (1973) noted that floods during warm seasons cause silting and an influx of excessive nutrients and organic matter, with consequent algal blooms and stratification of waters. These factors may, in turn, lead to low dissolved oxygen conditions. During other seasons, increased accumulations of detritus caused by increased riverflow and salinity alterations could actually be beneficial to microorganisms and detritivores such as isopods, amphipods, and some decapods. Detritus also serves as the major energy base utilized by juveniles of most fish species from sea grass beds (Carr and Adams 1973) and is probably important as a direct or indirect source of food for many fishes in the Santee system.

Sustained abundance of Penaeus setiferus in the Santee system is questionable following rediversion. Shrimp are known detritivores, and large areas of brackish/salt marsh and estuary with substantial land runoff are considered to be conducive to good shrimp production (Bishop and Shealy 1977). Rediversion will cause waters to inundate many areas and should result in a seaward progression of freshwater and brackish water plant communities. Because the total area of estuarine habitat should effectively be moved seaward, a decrease in actual acreage available as nursery habitat may result; yet lower salinity conditions are still likely to exert the greatest influence on shrimp production. Young P. setiferus are most abundant in salinities <10%, whereas young *P. aztecus* are most abundant in salinities from 10 to 20 % (Gunter et al. 1964). Despite these optimum ranges, Barrett and Gillespie (1973, 1975) have suggested that an inverse relationship exists between the amount of freshwater introduced into coastal Louisiana and the catches of brown and white shrimp. Also, increased turbidity and hypertrophy may inhibit photosynthesis so that an initial reduction in oxygen may occur in bottom waters. Others (Hildebrand and Gunter 1953; Copeland 1966; Aleem 1972; Glaister 1978) have noted a positive relationship between shrimp abundance and river discharge, but not all of these studies indicated that abundance was increased within the estuary.

Blue crabs will probably be little affected by rediversion because of their high mobility and tolerance of low-salinity conditions, but increased siltation from rediversion could hamper their respiration. Although blue crab populations sustained little damage following Hurricane Agnes, mortalities in Chesapeake Bay were attributed to increases siltation, low dissolved oxygen levels, and red tide (Chesapeake Bay Research Council 1973).

³Van Engel, W. A., and E. B. Joseph. 1968. Characterization of coastal and estuarine fish nursery grounds as natural communities. U.S. Fish Wildl. Serv. Final Rep., 43 p.

From the available data and published literature, it appears that abundance of resident species of decapod crustaceans and fishes from the Santee system will be enhanced by rediversion if riverflow increase is gradual and properly regulated during natural freshets and warm weather periods. Such regulation will insure that salinities do not reach levels below tolerance and that hypertrophic conditions do not occur. On the other hand, the effect of rediversion on transient species such as sciaenid fishes and penaeid shrimps may not be beneficial. A decrease in nursery habitat following rediversion would lower abundance of these species. This is indicated by two aspects of our results: Both biomass and density of the South Santee River, which currently receives less freshwater input, were higher than that of the North Santee River; and both abundance and biomass of dominant species appear to be generally lower at the stations furthest upriver. Species diversity will undoubtedly decrease due to decreased utilization of the lower portion of the Santee Rivers by marine stenohaline species. Lower salinity conditions at and near the mouth should deter penetration of the estuary by these species. Whether decreased abundance of marine transients in the vicinity of the upriver stations following rediversion will be offset by more optimum salinity conditions and increased abundance nearer the mouths of the rivers is supposition. It appears that rediversion is certain; therefore, it is imperative that careful monitoring of biological and hydrographic conditions occur during and after rediversion in order to ascertain effects on the estuarine biota.

ACKNOWLEDGMENTS

We are grateful to all persons who participated in collection of specimens and data in the field and workup of samples at the laboratory. We thank E. B. Joseph and V. G. Burrell, Jr., for their management support and advice during the field study; and Captain V. Taylor of the *RV Anita* and A. S. Lachicotte for vessel operations. The figures were drafted by K. Swanson and B. Stone; N. Kopacka skillfully created most of the computer programs used in data analysis; W. Coon helped with preparation of tables and proofed the manuscript, and B. Ashby typed the manuscript. We appreciate their patience and organization. The manuscript was improved through reviews by G. Sedberry, R. Van Dolah, C. A. Wenner, C. Bearden, and the two anonymous reviewers.

The data were collected and automated by support from Coastal Plains Regional Commission (Contract 10340031) as part of the Estuarine Survey Program of the South Carolina Marine Resources Center. First author's support and analysis of the data were funded by NOAA Office of Sea Grant (Grant 00140-93027).

LITERATURE CITED

- ALDERDICE, D. F.
- 1972. Factor combinations: Responses of marine poikilotherms to environmental factors acting in concert. *In* O. Kinne (editor), Marine ecology – a comprehensive, integrated treatise on life in oceans and coastal waters, Vol. 1, Part 3, p. 1659-1722. Wiley-Interscience, N.Y. ALEEM, A. A.
- 1972. Effect of river outflow management on marine life. Mar. Biol. (Berl.) 15:200-208.

ANDREWS, J. D.

1973. Effects of tropical storm Agnes on epifaunal invertebrates in Virginia estuaries. Chesapeake Sci. 14:223-234.

BARNES, H., and T. B. BAGENAL.

1951. A statistical study of variability in catch obtained by short repeated

trawls taken over an inshore ground. J. Mar. Biol. Assoc. U.K. 29:649-660.

BARRETT, B. B., and M. C. GILLESPIE.

 Primary factors which influence commercial shrimp production in coastal Louisiana. La. Wildl. Fish. Comm., Tech. Bull. 9, 28 p.
1975. 1975 environmental conditions relative to shrimp production in coastal

Louisiana. La. Wildl. Fish. Comm., Tech. Bull. 15, 22 p.

BISHOP, J. M., and M. H. SHEALY, Jr.

1977. Biological observations on commercial penaeid shrimps caught by bottom trawl in South Carolina estuaries, February 1973-January 1975. S.C. Mar. Resour. Cent. Tech. Rep. 25, 97 p.

BOESCH, D. F.

1977. A new look at the zonation of benthos along the estuarine gradient. In B. C. Coull (editor), Ecology of marine benthos, p. 245-266. Univ. South Carolina Press, Columbia.

BOESCH, D. F., R. J. DIAZ, and R. W. VIRNSTEIN.

1976. Effects of tropical storm Agnes on soft-bottom macrobenthic communities of the James and York estuaries and the lower Chesapeake Bay. Chesapeake Sci. 17:246-259.

BURRELL, V. G., Jr.

1977. Mortalities of oysters and hard clams associated with heavy runoff in the Santee River system, South Carolina in the Spring of 1975. Proc. Natl. Shellfish. Assoc. 67:35-43.

CALDER, D. R., B. B. BOOTHE, Jr., and M. S. MACLIN.

1977. A preliminary report on estuarine macrobenthos of the Edisto and Santee River systems, South Carolina. S.C. Mar. Resour. Cent. Tech. Rep. 22, 50 p.

CARR, W. E. S., and C. A. ADAMS.

- 1973. Food habits of juvenile marine fishes occupying seagrass beds in the estuarine zone near Crystal River, Florida. Trans. Am. Fish. Soc. 102:511-540.
- CHAO, L. N., and J. A. MUSICK.
 - 1977. Life history, feeding habits, and functional morphology of juvenile sciaenid fishes in the York River estuary, Virginia. Fish. Bull., U.S. 75:657-702.

CHESAPEAKE BAY RESEARCH COUNCIL.

1973. Effects of Hurricane Agnes on the environment and organisms of Chesapeake Bay: Early findings and recommendations. Va. Inst. Mar. Sci. Spec. Sci. Rep. Mar. Ocean Eng. 29, 172 p.

CLIFFORD, H. T., and W. STEPHENSON.

- 1975. An introduction to numerical classification. Acad. Press, N.Y., 229 p.
- COPELAND, B. J.

1966. Effects of decreased river flow on estuarine ecology. J. Water Pollut. Cont. Fed. 38:1831-1839.

COPELAND, B. J., and T. J. BECHTEL.

1974. Some environmental limits of six Gulf coast estuarine organisms. Contrib. Mar. Sci. 18:169-204.

DOVEL, W. L., J. A. MIHURSKY, and A. J. MCERLEAN.

1969. Life history aspects of the hogchocker, *Trinectes maculatus*, in the Patuxent River estuary, Maryland. Chesapeake Sci. 10:104-119.

ELLIOTT, J. M.

1977. Some methods for the statistical analysis of samples of benthic invertebrates. Freshwater Biol. Assoc., Sci. Publ. 25, 160 p.

GLAISTER, J. P.

- 1978. The impact of river discharge on distribution and production of the school prawn *Metapenaeus macleayi* (Haswell) (Crustacea: Penaeidae) in the Clarence River region, Northern New South Wales. Aust. J. Mar. Freshwater Res. 29:311-323.
- GUNTER, G.
 - 1938. Seasonal variations in abundance of certain estuarine and marine fishes in Louisiana, with particular reference to life histories. Ecol. Monogr. 8:313-346.
 - 1945. Studies on marine fishes of Texas. Publ. Inst. Mar. Sci. Univ. Tex. 1:1-190.
 - 1961. Some relations of estuarine organisms to salinity. Limnol. Oceanogr. 6:182-190.

GUNTER, G., J. Y. CHRISTMAS, and R. KILLEBREW.

1964. Some relations of salinity to population distributions of motile estuarine organisms with special reference to penaeid shrimp. Ecology 45:181-185.

GUNTER, G., and H. H. HILDEBRAND.

1951. Destruction of fishes and other organisms on the South Texas coast by the cold wave of January 28-February 3, 1951. Ecology 32:731-736. HAEDRICH, R. L., and S. O. HAEDRICH.

1974. A seasonal survey of the fishes in the Mystic River, a polluted

estuary in downtown Boston, Massachusetts. Estuarine Coastal Mar. Sci. 2:59-73.

HANSEN, D. J. 1969. Food, growth, migration, reproduction, and abundance of pinfish, Lagodon rhomboides, and Atlantic croaker, Micropogon undulatus, near Pensacola, Florida, 1963-65. Fish. Bull., U.S. 68:135-146.

HAVEN, D. S.

1957. Distribution, growth and availability of juvenile croaker, *Micropogon undulatus*, in Virginia. Ecology 38:88-97.

HILDEBRAND, H. H., and G. GUNTER.

1953. Correlation of rainfall with Texas catch of white shrimp, *Penaeus setiferus* (Linnaeus). Trans. Am. Fish. Soc. 82:151-155.

1930. Development and life history of fourteen teleostean fishes at Beaufort, N.C. Bull. U.S. Bur. Fish. 46:383-488.

1938. Further notes on the development and life history of some teleosts at Beaufort, N.C. Bull. U.S. Bur. Fish. 48:505-642.

HOESE, H. D.

1965. Spawning of marine fishes in the Port Aransas, Texas area as determined by the distribution of young and larvae. Ph.D. Dissertation, Univ. Texas, Austin, 150 p.

1973. A trawl study of nearshore fishes and invertebrates of the Georgia coast. Contrib. Mar. Sci. 17:63-98.

HOFF, J. G., and R. M. IBARA.

1977. Factors affecting the seasonal abundance, composition and diversity of fishes in a southeastern New England estuary. Estuarine Coastal Mar. Sci. 5:665-678.

KILBY, J. D.

1955. The fishes of two Gulf coastal marsh areas of Florida. Tulane Stud. Zool. 2(8):175-247.

KINNE, O.

1963. The effects of temperature and salinity on marine and brackish water animals. I. Temperature. Oceanogr. Mar. Biol., Annu. Rev. 1:301-340.

1966. Physiological aspects of animal life in estuaries with special reference to salinity. Neth. J. Sea Res. 3:222-244.

KJERFVE, B.

1976. The Santee-Cooper: a study of estuarine manipulations. In M. Wiley (editor), Estuarine processes, Vol. I, p. 44-56. Acad. Press, N.Y. KJERFVE, B., and J. E. GREER.

1978. Hydrography of the Santee River during moderate discharge conditions. Estuaries 1:111-119.

LAMBERT, J. M., and W. T. WILLIAMS.

1962. Multivariate methods in plant ecology. IV. Nodal analysis. J. Ecol. 50:775-802.

1967. A general theory of classificatory sorting strategies. I. Hierarchical systems. Comput. J. (Lond.) 9:373-380.

LIVINGSTON, R. J.

1976. Diurnal and seasonal fluctuations of organisms in a North Flordia estuary. Estuarine Coastal Mar. Sci. 4:373-400.

- LIVINGSTON, R. J., P. S. SHERIDAN, B. G. McLANE, F. G. LEWIS III, and G. G. KOBYLINSKI.
 - 1977. The biota of the Apalachicola Bay System: functional relationships. *In* R. J. Livingston and E. A. Joyce, Jr. (editors), Proceedings of the Conference on the Apalachicola Drainage System, 23-24 April, 1976, Gainesville, Fla. Fla. Mar. Res. Publ. 26, 177 p.

MANSUETI, A. J., and J. D. HARDY, Jr.

1967. Development of fishes of the Chesapeake Bay region. An atlas of egg, larval, and juvenile stages, Part 1. Nat. Res. Inst., Univ. Maryland, 202 p.

MARKLE, D. F.

1976. The seasonality of availability and movements of fishes in the channel of the York River, Virginia. Chesapeake Sci. 17:50-55.

MILLS, E. L.

1975. Benthic organisms and the structure of marine ecosystems. J. Fish. Res. Board Can. 32:1657-1663.

MUSICK, J. A., and J. D. McEACHRAN.

1972. Autumn and winter occurrence of decapod crustaceans in Chesapeake Bight, U.S.A. Crustaceana 22:190-200.

NELSON, F. P. (editor).

1976. Lower Santee River environmental quality study. S.C. Water Resour. Comm., Rep. 120, 60 p.

OVIATT, C. A., and S. W. NIXON.

1973. The demersal fish of Narragansett Bay: an analysis of community structure, distribution and abundance. Estuarine Coastal Mar. Sci. 1:361-378.

PEARSE, A. S.

1936. The migrations of animals from sea to land. Duke Univ. Press, Durham, N.C., 176 p.

REID, G. K., Jr.

1954. An ecological study of the Gulf of Mexico fishes, in the vicinity of Cedar Key, Florida. Bull. Mar. Sci. Gulf. Caribb. 4:1-94.

ROE, R. B.

1969. Distribution of royal-red shrimp, *Hymenopenaeus robustus*, on three potential commercial grounds off the southeastern United States. U.S. Fish Wildl. Serv., Fish. Ind. Res. FINR-A 5:161-174.

RUELLO, N. V.

1973. The influence of rainfall on the distribution and abundance of the school prawn *Metapenaeus macleayi* in the Hunter River Region (Australia). Mar. Biol. (Berl.) 23:221-228.

SHEALY, M. H., Jr., and J. M. BISHOP.

1979. Hydrographic and biological studies in relation to Cooper River low-flow conditions prior to river rediversion. *In* Cooper River controlled low-flow study, p. 139-178. S.C. Water Resour. Comm., Rep. 131.

SHEALY, M. H., Jr., J. V. MIGLARESE, and E. B. JOSEPH. 1974. Bottom fishes of South Carolina estuaries, relative abundance, seasonal distribution, and length-frequency relationships. S.C. Mar. Resour. Cent., Tech. Rep. 6, 189 p.

SOKAL, R. R., and F. J. ROHLF.

1969. Biometry. The principles and practice of statistics in biological research. W. H. Freeman and Co., San Francisco, 776 p.

STEPHENS, D. G., D. S. VAN NIEUWENHUISE, W. H. KANES, and T. T. DAVIES.

1975. Environmental analysis of the Santee River estuaries: thirty years after diversion. Southeast. Geol. 16:131-144.

STEVENS, R. E.

1959. The white and channel catfishes of the Santee-Cooper reservoir and tailrace sanctuary. 13th Annu. Conf. Southeast. Game Fish. Comm. p. 203-219.

STRICKLAND, J. D. H., and T. R. PARSONS.

1968. A practical handbook of seawater analysis. Fish. Res. Board Can., Bull. 167, 311 p.

SYMPOSIUM ON THE CLASSIFICATION OF BRACKISH WATERS.

1958. The Venice System for the classification of marine waters according to salinity. Oikos 9:311-312.

TAYLOR, C. C.

TURNER, W. R., and G. N. JOHNSON.

1974. Standing crops of aquatic organisms in tidal streams of the lower Cooper River system, South Carolina. *In* F. P. Nelson (editor), The Cooper River environmental study, p. 13-20. S.C. Water Resour. Comm., Rep. 117.

WALLACE, J. H., and R. P. VAN der ELST.

1975. The estuarine fishes of the east coast of South Africa. V. Ecology, estuarine dependence and status. S. Afr. Assoc. Mar. Biol. Res., Invest. Rep. 42:19-27.

WATHNE, F.

1959. Observations on trawl-door spread and a discussion of influencing factors. Commer. Fish. Rev. 21(10):7-15.

WEINSTEIN, M. P.

1979. Shallow marsh habitats as primary nurseries for fishes and shellfish, Cape Fear River, North Carolina. Fish. Bull., U.S. 77:339-357.

WHITE, M. L., and M. E. CHITTENDEN.

1976. Aspects of the life history of the Atlantic croaker, *Micropogon undulatus*. Texas A&M Sea Grant Publ. TAMU-GS-76-205, 54 p.

WILLIAMS, W. T., and J. M. LAMBERT. 1961. Multivariate methods in plant ecology. III. Inverse associationanalysis. J. Ecol. 49:717-729.

HILDEBRAND, S. F., and L. E. CABLE.

LANCE, G. N., and W. T. WILLIAMS.

^{1953.} Nature of variability in trawl catches. U.S. Fish Wildl. Serv., Fish. Bull. 54:145-166.